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54) Title: A METHOD OF DESIGNING ALPHA-AMYLASE MUTANTS WITH PREDETERMINED PROPERTIES

(57) Abstract

A method of constructing a variant of a paent Termamyl-like a-amylase, which variant has \(\alpha \)-amylase activity and at least one altered property as compared to the parent \(a \)-amylase, comprises i) analysing the structure of the parent Termamyl-like \(a \)-amylase is identify at least one attrion sciol residue or at least one attrion. The structure of the parent Termamyl-like \(a \)-amylase structure, which amino acid residue or at structural part of the Termamyl-like \(a \)-amylase structure, which amino acid residue or structural part is believed to be of relevance for altering the property of the parent Termamyl-like \(a \)-amylase (as evaluated on the said of structural part is described as compared to the parent Termamyl-like \(a \)-amylase, has been modified in the amino acid residue or structural part identified in \(j \) so as to alter the property, and iii) testing the resulting Termamyl-like \(a \)-amylase variant, which a camylase variant for the property in question.

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A METHOD OF DESIGNING ALPHA-AMYLASE MUTANTS WITH PREDETERMINED PROPERTIES

FIELD OF THE INVENTION

5 The present invention relates to a novel method of designing α -amylase mutants with predetermined properties, which method is based on the hitherto unknown three-dimensional structure of bacterial α -amylases.

10 BACKGROUND OF THE INVENTION

 α -Amylases (α -1,4 glucan-4-glucanohydrolase, EC 3.2.1.1) constitute a group of enzymes which is capable of hydrolyzing starch and other linear and branched 1,4-glucosidic oligo- and 15 polysaccharides. Almost all α -amylases studied have a few conserved regions with approximately the same length and spacing. One of these regions resembles the Ca2+ binding site of calmodulin and the others are thought to be necessary for the active centre and/or binding of the substrate.

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While the amino acid sequence and thus primary structure of a large number of α -amylases are known, it has proved very difficult to determine the three-dimensional structure of all α -amylases. The three-dimensional structure can be determined by X-ray crystallographic analysis of α -amylase crystals, but it has proven difficult to obtain α -amylase crystals suitable for actually solving the structure.

Until now the three-dimensional structure of only a few 30 α -amylases have been determined at high resolution. These include the structure of the Aspergillus oryzae TAKA α -amylase (Swift et al., 1991), the Aspergillus niger acid amylase (Brady et al, 1991), the structure of pig pancreatic α -amylase (Qian et al., 1993), and the barley alpha-amylase (Kadziola et al. 1994, Journal of Molecular Biology 239: 104-121, A.Kadziola, Thesis, Dept of Chemistry, U. of Copenhagen, Denmark). Furthermore, the three-dimensional structure of a Bacillus circulans cyclodextrin glycosyltransferase (CGTase) is known

(Klein et al., 1992) (Lawson et al., 1994). The CGTase catalyzes the same type of reactions as α -amylases and exhibits some structural resemblance with α -amylases.

5 Furthermore, crystallization and preliminary X-ray studies of B. subtilis α-amylases have been described (Chang et al. (1992) and Mizuno et al. (1993)). No final B. subtilis structure has been reported. Analogously, the preparation of B. licheniformis α-amylase crystals has been reported (Suzuki et al. (1990), but no subsequent report on X-ray crystallographic analysis or three-dimensional structure are available.

Several research teams have attempted build to three-dimensional structures on the basis of the above known 15 α-amylase structures. For instance, Vihinen et al. (J. Biochem. 107, 267-272, 1990), disclose the modelling (or computer simulation) of a three-dimensional structure of the Bacillus stearothermophilus \u03c3-amvlase on the basis of the TAKA amvlase structure. The model was used to investigate hypothetical 20 structural consequences of various site-directed mutations of the B. stearothermophilus α-amylase. E.A. MacGregor (1987) predicts the presence of α -helices and β -barrels in α -amylases from different sources, including barley, pig pancreas and Bacillus amyloliquefaciens on the basis of the known structure 25 of the A. oryzae TAKA α-amylase and secondary structure predicting algorithms. Furthermore, the possible loops and subsites which may be found to be present in, e.g., the B. amyloliquefaciens α -amylase are predicted (based on a comparison with the A. oryzae sequence and structure).

A.E. MacGregor (Starch/Stärke 45 (1993), No. 7, p. 232-237) presents a review of the relationship between the structure and activity of α -amylase related enzymes.

35 Hitherto, no three-dimensional structure has been available for the industrially important Bacillus α-amylases (which in the present context are termed "Termamyl-like α-amylases").

including the B. licheniformis, the B. amyloliquefaciens, and the B. stearothermophilus α -amylase.

BRIEF DISCLOSURE OF THE INVENTION

The three-dimensional structure of a Termamyl-like bacterial α -amylase has now been elucidated. On the basis of an analysis of said structure it is possible to identify structural parts or specific amino acid residues which from structural or 10 functional considerations appear to be important for conferring the various properties to the Termamyl-like α -amylases. Furthermore, when comparing the Termamyl-like α -amylase structure with known structures of the fungal and mammalian α -amylases mentioned above, it has been found that some 15 similarities exist between the structures, but also that some striking, and not previously predicted structural differences between the α -amylases exist. The present invention is based on these findings.

20 Accordingly, in a first aspect the invention relates to a method of constructing a variant of a parent Termamyl-like α -amylase, which variant has α -amylase activity and at least one altered property as compared to said parent α -amylase, which method comprises

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- i) analysing the structure of the Termamyl-like α -amylase with a view to identifying at least one amino acid residue or at least one structural part of the Termamyl-like α -amylase structure, which amino acid residue or structural part is 30 believed to be of relevance for altering said property of the parent Termamyl-like α -amylase (as evaluated on the basis of structural or functional considerations),
- ii) constructing a Termamyl-like α-amylase variant, which as 35 compared to the parent Termamyl-like α-amylase, has been modified in the amino acid residue or structural part identified in i) so as to alter said property, and

- iii) testing the resulting Termamyl-like α -amylase variant for said property.
- 5 In a second aspect the present invention relates to a method of constructing a variant of a parent Termamyl-like α -amylase, which variant has α -amylase activity and one or more altered properties as compared to said parent α -amylase, which method comprises
- 10 i) comparing the three-dimensional structure of the Termamyllike α -amylase with the structure of a non-Termamyllike α -amylase.
- ii) identifying a part of the Termamyl-like α -amylase structure which is different from the non-Termamyl-like α -amylase 15 structure, and
 - iii) modifying the part of the Termamyl-like α -amylase identified in ii) whereby a Termamyl-like α -amylase variant is obtained, one or more properties of which differ from the parent Termamyl-like α -amylase.

- In a third aspect the invention relates to a method of constructing a variant of a parent non-Termamyl-like α -amylase, which variant has α -amylase activity and one or more altered properties as compared to said parent α -amylase, which method 25 comprises
 - i) comparing the three-dimensional structure of the non-Termamyl-like α -amylase with the structure of a Termamyl-like α -amylase.
- ii) identifying a part of the non-Termamyl-like α -amylase structure which is different from the Termamyl-like α -amylase structure, and
- iii) modifying the part of the non-Termamyl-like α -amylase identified in ii) whereby a non-Termamyl-like α -amylase variant is obtained, one or more properties of which differ from the parent Termamyl-like α -amylase.

The property which may be altered by the above methods of the present invention may, e.g., be substrate specificity,

substrate binding, substrate cleavage pattern, temperature stability, pH dependent activity, pH dependent stability (especially increased stability at low (e.g. pH<6, in particular pH<5) or high (e.g. pH>9) pH values), stability 5 towards oxidation, Ca^{2*}-dependency, specific activity, and other properties of interest. For instance, the alteration may result in a variant which, as compared to the parent Termamyl-like α amylase, has an increased specific activity at a given pH and/or an altered substrate specificity.

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In still further aspects the invention relates to variants of a Termamyl-like α -amylase, DNA encoding such variants and methods of preparing the variants. Finally, the invention relates to the use of the variants for various industrial 15 purposes.

DETAILED DISCLOSURE OF THE INVENTION

The Termamyl-like \alpha-amylase

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and

It is well known that a number of alpha-amylases produced by Bacillus spp. are highly homologous on the amino acid level. For instance, the B. licheniformis α -amylase comprising the amino acid sequence shown in SEQ ID No. 2 (commercially 25 available as Termamyl®) has been found to be about 89% homologous with the B. amyloliquefaciens α-amylase comprising the amino acid sequence shown in SEO ID No. 4 and about 79% homologous with the B. stearothermophilus α -amylase comprising the amino acid sequence shown in SEO ID No. 6. Further 30 homologous α -amylases include an α -amylase derived from a strain of the Bacillus sp. NCIB 12289, NCIB 12512, NCIB 12513 or DSM 9375, all of which are described in detail in WC 95/26397, and the α -amylase described by Tsukamoto et al., 1988, Biochemical and Biophysical Research Communications, Vol. 35 151. No. 1. Still other homologous α -amylases include the α -

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amylase produced by the B. licheniformis described in EP 252 666 (ATCC 27811), and the α -amylases identified in WO 91/00353 WO 94/18314. Other commercial Termamyl-like E.

licheniformis α -amylases are Optitherm® and Takatherm® (available from Solvay), Maxamyl® (available from Gistbrocades/Genencor), Spezym AA® (available from Genencor), and Keistase® (available from Daiwa).

Because of the substantial homology found between these α -amylases, they are considered to belong to the same class of α -amylases, namely the class of "Termamyl-like α -amylases".

10 Accordingly, in the present context, the term "Termamyl-like α amylase" is intended to indicate an α -amylase which, on the amino acid level, exhibits a substantial homology to Termamyl®, i.e. the B. licheniformis α -amylase SEQ ID NO 2. In other words, a Termamyl-like α -amylase is an α -amylase, which has the 15 amino acid sequence shown in SEQ ID No. 2, 4 or 6 herein, or the amino acid sequence shown in SEQ ID NO 1 or 2 of WO 95/26397 or in Tsukamoto et al., 1988, or i) which displays at least 60%, such as at least 70%, e.g. at least 75%, or at least 80%, e.g. at least 85%, at least 90% or at least 95% homology 20 with at least one of said amino acid sequences and/or ii) displays immunological cross-reactivity with an antibody raised against at least one of said α -amylases, and/or iii) is encoded by a DNA sequence which hybridizes to the DNA sequences encoding the above specified \alpha-amylases which are apparent from 25 SEQ ID Nos. 1, 3 and 5 of the present application, and SEO ID NO 4 and 5 of WO 95/26397, respectively.

In connection with property i) the "homology" may be determined by use of any conventional algorithm, preferably by use of the 30 GAP progamme from the GCG package version 7.3 (June 1993) using default values for GAP penalties (Genetic Computer Group (1991) Programme Manual for the GCG Package, version 7, 575 Science Drive, Madison, Wisconsin, USA 53711).

35 Property ii) of the α -amylase, i.e. the immunological cross reactivity, may be assayed using an antibody raised against or reactive with at least one epitope of the relevant Termamyllike α -amylase. The antibody, which may either be monoclonal or

polyclonal, may be produced by methods known in the art, e.g. as described by Hudson et al., 1989. The immunological cross-reactivity may be determined using assays known in the art, examples of which are Western Blotting or radial immunodiffusion assay, e.g. as described by Hudson et al., 1989. In this respect, immunological cross-reactivity between the α -amylases having the amino acid sequences SEQ ID Nos. 2, 4 and 6, respectively, has been found.

The oligonucleotide probe used in the characterization of the Termamy1-like α-amylase in accordance with property iii) above may suitably be prepared on the basis of the full or partial nucleotide or amino acid sequence of the α-amylase in question. Suitable conditions for testing hybridization involve presoaking in 5xSSC and prehybridizing for 1h at -40°C in a solution of 20% formamide, 5xDenhardt's solution, 50mM sodium phosphate, pH 6.8, and 50μg of denatured sonicated calf thymus DNA, followed by hybridization in the same solution supplemented with 100μM ATP for 18h at -40°C, or other methods described by e.g. Sambrook et al., 1989.

In the present context, "derived from" is intended not only to indicate an α -amylase produced or producible by a strain of the organism in question, but also an α -amylase encoded by a DNA sequence isolated from such strain and produced in a host organism transformed with said DNA sequence. Finally, the term is intended to indicate an α -amylase which is encoded by a DNA sequence of synthetic and/or cDNA origin and which has the identifying characteristics of the α -amylase in question. The term is also intended to indicate that the parent α -amylase may be a variant of a naturally occurring α -amylase, i.e. a variant which is the result of a modification (insertion, substitution, deletion) of one or more amino acid residues of the naturally occurring α -amylase.

Parent hybrid a-amylases

The parent α -amylase (being a Termamyl-like or non-Termamyl-like α -amylase) may be a hybrid α -amylase, i.e. an α -amylase s which comprises a combination of partial amino acid sequences derived from at least two α -amylases.

The parent hybrid α -amylase may be one which on the basis of amino acid homology and/or immunological cross-reactivity and/or DNA hybridization (as defined above) can be determined to belong to the Termamyl-like α -amylase family. In this case, the hybrid α -amylase is typically composed of at least one part of a Termamyl-like α -amylase and part(s) of one or more other α -amylases selected from Termamyl-like α -amylases or non-15 Termamyl-like α -amylases of microbial (bacterial or fungal) and/or mammalian origin.

Thus, the parent hybrid α -amylase may comprise a combination of at least two Termamyl-like α -amylases, or of at least one 20 Termamyl-like and at least one non-Termamyl-like bacterial α amylase, or of at least one Termamyl-like and at least one fungal α -amylase. For instance, the parent α -amylase comprises a C-terminal part of an α -amylase derived from a strain of B. licheniformis and a N-terminal part of an α -amylase derived 25 from a strain of B. amyloliquefaciens or from a strain of B. stearothermophilus. For instance, the parent comprises at least 430 amino acid residues of the C-terminal part of the B. licheniformis α-amylase, and may, e.g. comprise a) an amino acid segment corresponding to the 37 N-terminal 30 amino acid residues of the B. amyloliquefaciens α -amylase having the amino acid sequence shown in SEQ ID No. 4 and an amino acid segment corresponding to the 445 C-terminal amino acid residues of the B. licheniformis \alpha-amylase having the amino acid sequence shown in SEQ ID No. 2, or b) an amino acid 35 segment corresponding to the 68 N-terminal amino acid residues of the B. stearothermophilus α -amylase having the amino acid sequence shown in SEQ ID No. 6 and an amino acid segment corresponding to the 415 C-terminal amino acid residues of the

B. licheniformis α -amylase having the amino acid sequence shown in SEO ID No. 2.

Analogously, the parent hybrid α -amylase may belong to a non-5 Termamyl-like α -amylase family, e.g. the Fungamyl-like α -amylase family. In that case the hybrid may comprise at least one part of an α -amylase belonging to the non-Termamyl-like α -amylase family in combination with one or more parts derived from other α -amylases.

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The three-dimensional Termamyl-like α-amylase structure

The Termamyl-like α -amylase which was used to elucidate the three-dimensional structure forming the basis for the present invention consists of the 300 N-terminal amino acids of the B. amyloliquefaciens α -amylase (with the amino acid sequence shown in SEQ ID No. 4) and amino acids 301-483 of the C-terminal end of the B. licheniformis α -amylase with the amino acid sequence SEQ ID No. 2. The bacterial α -amylase belongs to the "Termamyl-like α -amylase family" and the present structure is believed to be representative for the structure of any Termamyl-like α -amylase.

The structure of the α -amylase was solved in accordance with 25 the principle for X-ray crystallographic methods given in "X-Ray Structure Determination", Stout, G.K. and Jensen, L.H., John Wiley & Sons, inc. NY, 1989. The structural coordinates for the solved crystal structure of the α -amylase at 2.2 Å resolution using the isomorphous replacement method are given 10 a standard PDB format (Brookhaven Protein Data Base) in Appendix 1. It is to be understood that Appendix 1 forms part of the present application.

Amino acid residues of the enzyme are identified by three-35 letter amino acid code (capitalized letters).

The α -amylase structure is made up of three globular domains ordered A, B, and C with respect to sequence, which lie

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approximately along a line in the order B, A, C. The domains can be defined as being residues 1-103 and 206-395 for domain A, residues 104-205 for domain B, and residues 396-483 for domain C, the numbers referring to the B. licheniformis α -5 amylase. This gives rise to an elongated molecule, the longest axis being about 85Å. The widest point perpendicular to this axis is approximately 50Å and spans the central A domain. The active site residues of the B. licheniformis α-amylase (SEQ ID NO 2) are D323. D231 and E261.

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Domain A

Domain A is the largest domain and contains the active site (comprised of a cluster of three amino acid residues placed at 15 the bottom of a deep cleft in the enzyme's surface). Domain A of all known α-amylase structures have the same overall fold, viz. the (beta/alpha)8 barrel with 8 central beta strands (number 1-8) and 8 flanking α -helices. The β -barrel is defined by McGregor op. cit. The C-terminal end of Beta strand 1 is 20 connected to helix 1 by a loop denoted loop 1 and an identical pattern is found for the other loops. These loops show some variation in size and some can be quite extensive.

The 8 central Beta-strands in the (beta/alpha)8 barrel 25 superimpose well between the various known α-amylase structures, and this part of the structure, including the close surroundings of the active site located at the c-terminal end of the beta-strands, show high similarity between the different amylases.

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The loops connecting beta-strands and alpha helices display high variations between alpha amylases. These loops constitute the structural context of the active site and the majority of the contacts to the substrate is found among residues located 35 in these loops. Such important characteristics as substrate specificity, substrate binding, pH/activity profile, starch cleavage pattern are determined by the amino acids and the positions of same in these loops.

The substantial differences between the Fungamyl-like α -amylase structure and the structure of the Termamyl-like α -amylase disclosed herein which are found in loops 1, 2, 3, and 8 are visualized in the Figures.

Domain B

The Termamyl-like α -amylase structure has been found to comprise a special domain structure in the λ domain's loop3, 10 also called domain B. The structure of the Termamyl-like α -amylase B domain has never been seen before in any of the known α -amylase or (β /alpha)8-barrel proteins.

The domain B structure is a very compact domain having a very 15 high number of charged residues. The B domain arises as an extension of the loop between strand 3 and helix 3 of domain A (shown in Fig. 7) and contains a 5 stranded antiparallel β -sheet structure containing at least one long loop structure and having the connectivity -1, +3, -1X, +2 (Richardson, 1981, Adv. 20 Protein Chem. 34, 167-339).

The first four strands of the B domain form two hairpin loops which twist around each other like a pair of crossed fingers (right-hand twist). The mainchain folds into a B-strand which 25 connects two small B-sheet structures. After making one turn in one sheet it folds back and makes up a two stranded sheet in contact with domain A and an internal hole in the α -amylase structure. Then the mainchain folds up to a small sheet structure nearly perpendicular to the first two sheets. Before entering the helix 3 on top of the B-strand 3, the approximately 24 last amino acids in domain B form two calcium binding sites in the contact region to domain A.

Domain B is connected with domain A by two peptide stretches, 35 which divide the domain-domain contact areas into two. Domain B is in contact with Domain A by a calcium binding region and an internally buried hole containing waters. Many types of molecular contacts are present. Ionic interacting between acid

and basic amino acids are possible, these interactions are very important for the general stability at high pH and for keeping the Calcium binding sites intact.

5 Domain C

Domain C is the C-terminal part of the protein consisting of amino acids 394-483. Domain C is composed entirely of β -strands which forms a single 8-stranded sheet structure, which folds 10 back on itself, and thus may be described as a β -sandwich structure. The connectivity is +1,+1, +5, -3, +1, +1, -3although strands 6 and 7 are only loosely connected. One part of the β -sheet forms the interface to domain A.

15 Ca-binding and Na-binding sites

The structure of the Termamyl-like α-amylase is remarkable in that it exhibits four calcium-binding sites and one sodiumbinding site. In other words four calcium ions and one sodium 20 ion are found to be present in the structure, although one of the calcium ions displays very weak coordination. Two of the calcium ions form part of a linear cluster of three ions, the central ion being attributed to sodium, which lie at the junction of the A and B domains.

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The coordinating residues for the calcium ions between the A and B domain are as follows (using the Pdb file nomenclature for amino acid residues and atoms in the Pdb file found in Appendix 1 herein): For the calcium ion nearest to the active 30 site (IUM 502 in the pdb file), the backbone carbonyls from His235 and Asp194, the sidechain atom OD1 from residues Asp194, Asn102 and Asp200, and one water molecule WAT X3 (atom OW7). For the sodium ion (IUM 505), the binding site includes atom OD2 from Asp194, Asp200, Asp183 and Asp159, and a backbone 35 carbonyl from Val201. The coordinates for the other calcium ion between domain A and B are (IUM 501) : atom OD2 from Asp204 and Asp159, backbone carbonyl from Asp183 and Ala181, atom OD1 from Asp202, and one water molecule WAT X7 (atom OW7).

One calcium ion is located between the A and C domain, another is located in the C domain. The first mentioned calcium ion, which is also the one best coordinated (IUM 503) includes a carbonyl backbone from Gly300, Tyr302 and His406, atom OD2/OD1 5 from Asp430, atom OD1 from Asp407, and one water molecule WAT X6 (atom OW7). The other and very weakly coordinated calcium site (IUM 504) comprises 4 water molecules WAT X21 (atom OW8), X6 (atom OW6), X9 (atom OW0) and X28 (atom OW8), OE1/OE2 from Glu447 and OD1 from Asp444

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Substrate-binding site

Without being limited to any theory it is presently believed that favourable interactions between a substrate molecule and 15 the enzyme (such as hydrogen bonds and/or strong electrostatic interaction) are found within a sphere of 4Å of the substrate, when bound to the enzyme. The following residues of the B. licheniformis α -amylase having the amino acid sequence shown in SEQ ID No. 2 are contemplated to be within a distance of 4 Å of the substrate and thus believed to be involved in interactions with the substrate:

Trp13, Tyr14, Asn17, Asp18, Ser50, Gln51, Ala52, Asp53, Val54, Gly55, Tyr56, Lys70, Arg74, Lys76, Val102, His105, Gly107, Gly108, Ala109, Trp138, Thr163, Asp164, Trp165, Asn172, Glu189, 25 Tyr193, Leu196, Met197, Tyr198, Ala199, Arg229, Asp231, Ala232, Lys234, His235, Glu261, Trp263, His327, Asp328, Gln333, Ser334, and Leu335.

The amino acid residues of another Termamyl-like α -amylase, which are contemplated to be within a distance of 4Å of the substrate, may easily be identified by aligning the amino acid sequence SEQ ID NO 2 with that of the other Termamyl-like α -amylase and thereby identifying the positions equivalent to those identified above.

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Generality of structure

Because of the high homology between the various Termamyl-like α -amylases, the solved structure defined by the coordinates of 5 Appendix 1 is believed to be representative for the structure of all Termamyl-like α -amylases. A model structure of other Termamyl-like α -amylases may easily be built on the basis of the coordinates given in Appendix 1 adapted to the α -amylase in question by use of an alignment between the respective amino 10 acid sequences. The creation of a model structure is exemplified in Example 1.

The above identified structurally characteristic parts of the Termamyl-like α -amylase structure (Ca-binding site, substrate 15 binding site, loops, etc.) may easily be identified in other Termamyl-like α -amylases on the basis of a model (or solved) structure of the relevant Termamyl-like α -amylase or simply on the basis of an alignment between the amino acid sequence of the Termamyl-like α -amylase in question with that of the B. 20 lichenformis α -amylase used herein for identifying the amino

acid residues of the respective structural elements.

Furthermore, in connection with Termamyl-like variants of the invention, which are defined by modification of specific amino acid residues of a specific Termamyl-like α -amylase, it will be understood that variants of another Termamyl-like α -amylase modified in an equivalent position (as determined from the best possible amino acid sequence alignment between the respective sequences) are intended to be covered as well. Thus, irrespective of whether an amino acid residue is identified herein for the purpose of defining a structural part of a given α -amylase or used for identifying a variant of the α -amylase, this amino acid residue shall be considered as representing the equivalent amino acid residue of any other Termamyl-like α -amylase.

Methods of the invention for design of novel α-amylase variants

In the methods according to the first, second and third aspects of the invention the terms "structure of a Termamyl-like α -5 amylase" and "Termamyl-like α -amylase structure" are intended to indicate the solved structure defined by the coordinates presented in Appendix 1 or a model structure of a given Termamyl-like α -amylase (such as the B. licheniformis α -amylase) built on the basis of the solved structure.

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In most cases the parent Termamyl-like α -amylase to be modified in accordance with the present invention is different from the α -amylase which was actually used for solving the structure (Appendix 1). This means that the amino acid residue(s) or 15 structural part(s) identified in the solved structure (Appendix 1) in step i) of the method according to the first, second or third aspect of the invention must be translated into the corresponding amino acid residue(s) or structural part(s) of the parent Termamyl-like α -amylase in question. 20 "translation" is conveniently performed on the basis of an amino acid sequence alignment between the amino acid sequence of the Termamyl-like α -amylase used for solving the structure and the amino acid sequence of the parent Termamyl-like α amylase in question.

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The analysis or comparison performed in step i) of the method according to the first, second and third aspect, respectively, of the invention may be performed by use of any suitable computer programme capable of analysing and/or comparing protein structures, e.g. the computer programme Insight, available from Biosym Technologies, Inc. For instance, the basic principle of structure comparison is that the three-dimensional structures to be compared are superimposed on the basis of an alignment of secondary structure elements (such as the central 8 \$\beta\$-strands in the barrel) and the parts differing between the structures can subsequently easily be identified from the superimposed structure.

The structural part which is identified in step i) of the methods of the first, second and third aspects of the invention may be composed of one amino acid residue. However, normally the structural part comprises more than one amino acid residue, typically constituting one of the above parts of the Termamyllike α -amylase structure such as one of the A, B, or C domains, an interface between any of these domains, a calcium binding site, a loop structure, the substrate binding site, or the like.

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In the present context the term "structural or functional considerations" is intended to indicate that modifications are made on the basis of an analysis of the relevant structure or structural part and its contemplated impact on the function of 15 the enzyme. Thus, an analysis of the structures of the various α -amylases, which until now has been elucidated, optionally in combination with an analysis of the functional differences between these α -amylases, may be used for assigning certain properties of the α -amylases to certain parts of the α -amylase 20 structure or to contemplate such relationship. For instance, differences in the pattern or structure of loops surrounding the active site may result in differences in access to the active site of the substrate and thus differences in substrate specificity and/or cleavage pattern. Furthermore, parts of a 25 Termamyl-like α-amylase involved in or contemplated to be involved substrate binding (and thus specificity/cleavage pattern), calcium or sodium ion binding (e.g. of importance for the Calcium-dependency of the enzyme), and the like has been identified (vide infra).

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The modification of an amino acid residue or structural part is typically accomplished by suitable modifications of a DNA sequence encoding the parent enzyme in question. The term "modified" as used in step ii) in the method according to the 35 first aspect of the invention is intended to have the following meaning: When used in relation to an amino acid residue the term is intended to mean replacement of the amino acid residue in guestion with another amino acid residue. When used in

relation to a structural part, the term is intended to mean replacement of one or more amino acid residues of said structural part, addition of one or more amino acid residues to said part, or deletion of one or more amino acid residues of said structural part.

The construction of the variant of interest is accomplished by cultivating a microorganism comprising a DNA sequence encoding the variant under conditions which are conducive for producing the variant, and optionally subsequently recovering the variant from the resulting culture broth. This is described in detail further below.

First aspect of the invention

15 In a preferred embodiment of the method according to the first aspect of the invention the property of the parent enzyme to be modified is selected from calcium dependency, substrate binding, cleavage pattern, pH dependent activity and the like. Specific examples of how to change these properties of a parent 20 Termamyl-like α-amylase are given further below.

In another preferred embodiment the parent Termamyl-like α -amylase to be modified is a *B. licheniformis* α -amylase.

25 Second and third aspects of the invention

One important advantage of the methods according to the second and third aspects of the present invention is that it is possible to adapt the structure (or a structural part) of a Termamyl-like α-amylase to the structure (or structural part)

30 of a non-Termamyl-like α-amylase and vide versa. For instance, having identified a loop structure of the non-Termamyl-like α-amylase which is believed to be responsible for or contributing to a particular property of the non-Termamyl-like α-amylase it is possible to replace the corresponding structure of the 35 Termamyl-like α-amylase with said non-Termamyl-like α-amylase structure - or if no corresponding structure exists in the Termamyl-like α-amylase - to insert the structure into the Termamyl-like α-amylase in such a manner that the resulting

variant Termamyl-like α -amylase, as far as the relevant part is concerned, resembles the corresponding part of the non-Termamyl-like α -amylase. When two or more parts of the structure of the parent Termamyl-like α -amylase are modified so as to resemble the corresponding parts of the non-Termamyl-like α -amylase it is possible to increase the resemblance to the non-Termamyl-like α -amylase of the Termamyl-like α -amylase variant and thus to alter the properties of said variant in the direction of those of said non-Termamyl-like α -amylase. Loop modifications are discussed in much further detail further below.

Typically, the modification to be performed in step iii) of the method according to the second aspect of the invention is accomplished by deleting one or more amino acid residues of the part of the Termamyl-like α -amylase to be modified so as to adapt the structure of said part of the parent α -amylase to the corresponding part of the non-Termamyl-like α -amylase; by replacing one or more amino acid residues of the part of the Termamyl-like α -amylase to be modified with the amino acid residues occupying corresponding positions in the non-Termamyl-like α -amylase; or by insertion of one or more amino acid residues present in the non-Termamyl-like α -amylase into a corresponding position in the Termamyl-like α -amylase. For the 2s method according to the third aspect the modification is to be understood analogously, performed on the non-Termamyl-like α -amylase rather than the Termamyl-like α -amylase.

In step ii) of the method according to the second or third
as aspect of the invention the part of the structure to be
identified is preferably one which in the folded enzyme is
believed to be in contact with the substrate (of the disclosure
above in the section entitled "Substrate-binding site) or
involved in substrate specificity and/or cleavage pattern,
sand/or one which is in contact with one of the calcium or
sodium ions and/or one, which is contributing to the pH or
temperature profile of the enzyme, or one which otherwise, from
structural or functional considerations, is contemplated to be

responsible for differences in one or more properties of the Termamyl-like and non-Termamyl-like α -amylase.

Non-Termamyl-like α -amylase

5 The non-Termamyl-like α -amylase with which the comparison is made in step i) of the method of the second aspect of the invention and which is the parent α -amylase in the method of the third aspect of the invention, may be any α -amylase, which does not belong to the family of Termamyl-like α -amylases (as defined above) and, which as a consequence thereof, has a different three-dimensional structure. Furthermore, the non-Termamyl-like α -amylase should be one which has, at the time that the method is performed, an elucidated or contemplated three-dimensional structure.

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The non-Termamyl-like α -amylase may, e.g., be a fungal α -amylase, a mammalian or a plant α -amylase or a bacterial α -amylase (different from a Termamyl-like α -amylase). Specific examples of such α -amylases include the Aspergillus oryzae TAKA 20 α -amylase, the A. niger acid α -amylase, the Bacillus subtilis α -amylase, the porcine pancreatic α -amylase and a barley α -amylase. All of these α -amylases have elucidated structures which are clearly different from the structure of the Termamyl-like α -amylase shown herein.

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The fungal α -amylases mentioned above, i.e. derived from A. niger and A. oryzae, are highly homologous on the amino acid level and generally considered to belong to the same family of α -amylases. In the present disclosure, this family is termed Trungamyl-like α -amylase and intends to indicate an α -amylase which exhibits a high homology, i.e. more than 70%, such as 80% homologous (as defined herein) to the fungal α -amylase derived from Aspergillus oryzae, commercially available as Fungamyl®, and the A. niger α -amylase.

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From the enclosed illustrations of the α -amylase structure of a Termamyl-like α -amylase and a comparison of said structure with the structure of a Fungamyl-like α -amylase it is evident

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that major differences exist between the two structures. In the method of the invention it is of particular interest to modify parts of the parent Termamyl-like α -amylase, which belong to a region with large differences to the Fungamyl-like α -amylase. 5 In particular, it is of interest to modify the parent Termamyllike α -amylase in one or more of the following loops: loop 1, loop 2, loop 3 and/or loop 8 of the parent α -amylase.

In the method of the third aspect of the invention it is of 10 particular interest to modify loop 1, loop 2, loop 3 and/or loop 8 of the parent non-Termamyl-like α -amylase to a closer ressemblance to the similar loops of a Termamyl-like α -amylase. such as Termamvl.

15 In the following specific types of variants are described which have been designed by use of the method of the invention.

Loop modifications

20 In order to change the substrate specificity of the parent α amylase to be modified it is relevant to consider loop modifications. For instance changing one or more of the loop structures of the Termamyl-like \alpha-amylase into a closer ressemblance with the corresponding loop structure(s) of a non-25 Termamyl-like α -amylase (such as a Fungamyl-like α -amylase) it is contemplated that it is possible to change the substrate specificity in the direction of that of the non-Termamyl α amylase. In the following different types of loop modifications of interest are listed. It will be understood that the variants 30 may have other changed properties in addition to the modified substrate specificity. It will be understood that the following modifications identified for a specific Termamyl-like α -amylase are intended to include corresponding modifications in other equivalent positions of other Termamyl-like α -amylases. 35 Furthermore, it will be understood that, normally, the loop

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modification will comprise replacement of an entire loop structure or a substantial part thereof in, e.g., the Termamyllike α -amylase, with the corresponding loop structure (or substantial part thereof) in a non-Termamyl-like α -amylase.

Loop2 modifications

- 5 In one embodiment the invention relates to a variant of a parent Termamyl-like α -amylase, in which variant at least one amino acid residue of the parent α -amylase, which is/are present in a fragment corresponding to the amino acid fragment 44-57 of the amino acid sequence of SEQ ID No. 4, i.e. loop 2,
- 10 has been deleted or replaced with one or more amino acid residues which is/are present in a fragment corresponding to the amino acid fragment 66-84 of the amino acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has been added using the relevant part of SEQ ID No.
- 15 10 or a corresponding part of another Fungamyl-like $\alpha\text{-amylase}$ as a template.

The amino acid sequence shown in SEQ ID No. 10 is the amino acid sequence of the A. cryzae α -amylase, i.e. a Fungamyl-like 20 α -amylase. It will be understood that amino acid residues or fragments found in corresponding positions in other α -amylases, in particular Fungamyl-like α -amylases, may be used as a template for the construction of the variant according to the invention. The corresponding part in other homologous α -25 amylases may easily be identified on the basis of a comparison of the amino acid sequences and/or three-dimensional structures of the respective α -amylases.

For instance, the variant may be one, which, when the amino acid sequence of the variant is aligned most closely with the amino acid sequence of the said parent a-amylase, occupies the same position as the portion from residue X to residue Y of SEQ ID No 4, the said region having at least 80% such as at least 90% sequence homology with the part of SEQ ID No 10 extending 35 from residue Z to residue V of SEQ ID No 10, wherein

X is the amino acid residue occupying position 44, 45, 46, 47 or 48 of SEQ ID No. 4,

Y is the amino acid residue occupying position 51, 52, 53, 54, 55, 56 or 57 of SEO ID No. 4,

- Z is the amino acid residue occupying position 66, 67, 68, 69 or 70 of SEQ ID No. 10, and
- 5 V is the amino acid residue occupying position 78, 79, 80, 81, 82, 83 or 84 of SEQ ID No. 10.

In other words, the variant may be one in which an amino acid fragment X-Y of the parent α -amylase, which corresponds to or 10 is within the amino acid fragment 44-57 of SEQ ID No. 4, has been replaced with an amino acid fragment Z-V, which corresponds to or is within the amino acid fragment 66-84 of the amino acid sequence shown in SEQ ID No. 10, in X, Y, Z and V have the meaning indicated above.

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A specific example of a variant according to this embodiment is a variant of a parent Termamyl-like α -amylase, in which the amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 48-51 of SEQ ID No. 4, has been replaced with an amino acid fragment corresponding to amino acid residues 70-78 of the amino acid sequence shown in SEQ ID No. 10.

Loop 3 modifications - limited alteration

25 In another embodiment the invention relates to a variant of a parent Termamyl-like \(\alpha \)-amylase, in which variant at least one of the amino acid residues of the parent \(\alpha \)-amylase, which is/are present in an amino acid fragment corresponding to the amino acid fragment 195-202 of the amino acid sequence of SEQ 30 ID No. 4, has been deleted or replaced with one or more of the amino acid residues which is/are present in an amino acid fragment corresponding to the amino acid fragment 165-177 of the amino acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has been added using the 35 relevant part of SEQ ID No. 10 or a corresponding part of another Fungamyl-like \(\alpha \)-amylase as a template.

For instance, the variant may be one in which an amino acid fragment X-Y of the parent α -amylase which corresponds to or is within the amino acid fragment 195-202 of SEQ ID No. 4, has been replaced by an amino acid fragment Z-V, which corresponds to or is within the amino acid fragment 165-177 of the amino acid sequence shown in SEQ ID No. 10, in which

X is an amino acid residue corresponding to the amino acid occupying position 195 or 196 of SEQ ID No. 4,

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Y is an amino acid residue corresponding to the amino acid occupying position 198, 199, 200, 201, or 202 of SEQ ID No. 4,

Z is an amino acid residue corresponding to the amino acid 15 occupying position 165 or 166 of SEQ ID No. 10, and

V is an amino acid residue corresponding to the amino acid occupying position 173, 174, 175, 176 or 177 of SEO ID No. 10.

- 20 Expressed in another manner, the variant according to this aspect may be one, which, when the amino acid sequence of variant is aligned most closely with the amino acid sequence of the said parent Termamyl-like α -amylase, occupies the same position as the portion from residue X to residue Y of SEQ ID
- 25 No 4, the said region having at least 80%, such as 90% sequence homology with the part of SEQ ID No 10 extending from residue Z to residue V of SEQ ID No 10, the meaning of X, Y, Z and V being as identified above.
- 30 A specific example of a variant according to this embodiment is a variant of a parent Termamyl-like α -amylase, in which the amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 196-198 of SEQ ID No. 4, has been replaced with the amino acid fragment corresponding to amino 35 acid residues 166-173 of the amino acid sequence shown in SEQ ID No. 10.

Loop 3 modifications - complete domain B

another Fungamyl-like α -amylase as a template.

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In a further embodiment the invention relates to a variant of a parent Termamyl-like α-amylase, in which variant at least one of the amino acid residues of the parent α -amylase, which 5 is/are present in a fragment corresponding to the amino acid

- fragment 117-185 of the amino acid sequence of SEO ID No. 4. has/have been deleted or replaced with one or more of the amino acid residues, which is/are present in an amino acid fragment corresponding to the amino acid fragment 98-210 of the amino 10 acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has been added using the relevant part of SEQ ID No. 10 or a corresponding part of
- 15 For instance, the variant may be one, in which an amino acid fragment X-Y of the parent α -amylase, which corresponds to or is within the amino acid fragment 117-185 of SEO ID No. 4, has been replaced with an amino acid fragment Z-V, which corresponds to or is within the amino acid fragment 98-210 of 20 the amino acid sequence shown in SEQ ID No. 10, in which variant
- X is an amino acid residue corresponding to the amino acid occupying position 117, 118, 119, 120 or 121 of SEQ ID No. 4, 25

Y is an amino acid residue corresponding to the amino acid occupying position 181, 182, 183, 184 or 185 of SEQ ID No. 4, Z is an amino acid residue corresponding to the amino acid occupying position 98, 99, 100, 101, 102 of SEQ ID No. 10, and

V is an amino acid residue corresponding to the amino acid occupying position 206, 207, 208, 209 or 210 of SEQ ID No. 10.

A specific example of a variant according to this embodiment is 35 a variant of a parent α -amylase, in which an amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 121-181 of SEQ ID No. 4, has been replaced with

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the amino acid fragment corresponding to amino acid residues 102-206 of the amino acid sequence shown in SEQ ID No. 10.

In another embodiment the invention relates to a variant of a 5 parent Termamyl-like α-amylase, in which variant at least one of the amino acid residues of the parent α-amylase, which is/are present in a fragment corresponding to the amino acid fragment 117-181 of the amino acid sequence of SEQ ID No. 4, has/have been deleted or replaced with one or more of the amino acid residues, which is/are present in an amino acid fragment corresponding to the amino acid fragment to 98-206 of the amino acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has been added using the relevant part of SEQ ID No. 10 or a corresponding part of 15 another Fungamyl-like α-amylase as a template.

For instance, the variant may be one, in which the amino acid fragment X-Y of the parent α -amylase, which corresponds to or is within the amino acid fragment 117-177 if SEQ ID No. 4, 20 has/have been replaced with an amino acid fragment Z-V, which corresponds to or is within the amino acid fragment 98-202 of the amino acid sequence shown in SEQ ID No. 10, in which variant

- 25 X is an amino acid residue corresponding to the amino acid occupying position 117, 118, 119, 120 or 121 of SEQ ID No. 4,
 - Y is an amino acid residue corresponding to the amino acid occupying position 174, 175, 176 or 177 of SEQ ID No. 4,
 - Z is an amino acid residue corresponding to the amino acid occupying position 98, 99, 100, 101, 102 of SEQ ID No. 10, and
- V is an amino acid residue corresponding to the amino acid $_{\rm 35}$ occupying position 199, 200, 201 or 202 of SEQ ID No. 10.

A specific example of a variant according to this embodiment of the invention is a variant, in which the amino acid fragment of

the parent α -amylase, which corresponds to amino acid residues 121-174 of SEQ ID No. 4, has been replaced with the amino acid fragment corresponding to amino acid residues 102-199 of the amino acid sequence shown in SEQ ID No. 10.

Loop 1 modifications - minimal addition

In a further embodiment the present invention relates to a variant of a parent Termamyl-like α-amylase, in which variant at least one of the amino acid residues of the parent α-10 amylase, which is/are present in an amino acid fragment corresponding to the amino acid fragment 12-19 of the amino acid sequence of SEQ ID No. 4, has/have been deleted or replaced with one or more of the amino acid residues, which is/are present in an amino acid fragment which corresponds to 15 the amino acid fragment 28-42 of SEQ ID No. 10, or in which one or more additional amino acid residues has/have been inserted using the relevant part of SEQ ID No. 10 or a corresponding part of another Fungamyl-like α-amylase as a template.

- 20 For instance, the variant may be one, in which the amino acid fragment X-Y of the parent α-amylase, which corresponds to or is within the amino acid fragment 12-19 of SEQ ID No. 4, has/have been replaced with an amino acid fragment Z-V, which corresponds to or is within the amino acid fragment 28-42 of the amino acid sequence shown in SEQ ID No. 10, in which variant.
 - $\rm X$ is an amino acid residue corresponding to the amino acid occupying position 12, 13 or 14 of SEQ ID No. 4,
 - Y is an amino acid residue corresponding to the amino acid occupying position 15, 16, 17, 18 or 19 of SEQ ID No. 4,
- Z is an amino acid residue corresponding to the amino acid 35 occupying position 28, 29, 30 , 31 or 32 of SEQ ID No. 10, and
 - V is an amino acid residue corresponding to the amino acid occupying position 38, 39, 40, 41 or 42 of SEQ ID No. 10.

A specific example of a variant according to this aspect of the invention is a variant, in which the amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 14-15 of SEQ ID No. 4, has been replaced with the amino acid 5 fragment corresponding to amino acid residues 32-38 of the amino acid sequence shown in SEQ ID No. 10.

Loop 1 modifications - complete loop

In a further embodiment the invention relates to a variant of a parent Termamyl-like α -amylase, in which variant at least one of the amino acid residues of the parent α -amylase, which is present in a fragment corresponding to amino acid residues 7-23 of the amino acid sequence of SEQ ID No. 4, has/have been deleted or replaced with one or more amino acid residues, which is/are present in an amino acid fragment corresponding to amino acid residues 13-45 of the amino acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has/have been inserted using the relevant part of SEQ ID No. 10 or a corresponding part of another Fungamyl-like α -amylase as a template.

For instance, the variant may be one, in which the amino acid fragment X-Y of the parent α -amylase, which corresponds to or is within the amino acid fragment 7-23 of SEQ ID No. 4, 25 has/have been replaced with an amino acid fragment Z-V, which corresponds to or is within the amino acid fragment 13-45 of the amino acid sequence shown in SEQ ID No. 10, in which variant

- 30 X is an amino acid residue corresponding to the amino acid occupying position 7 or 8 of SEQ ID No. 4,
 - Y is an amino acid residue corresponding to the amino acid occupying position 18, 19, 20, 21, 22 or 23 of SEQ ID No. 4,
 - Z is an amino acid residue corresponding to the amino acid occupying position 13 or 14 of SEQ ID No. 10, and

V is an amino acid residue corresponding to the amino acid occupying position 40, 41, 42, 43, 44 or 45 of SEQ ID No. 10.

A specific variant according to this embodiment is one, in s which the amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 8-18 of SEQ ID No. 4, has been replaced with the amino acid fragment corresponding to amino acid residues 14-40 of the amino acid sequence shown in SEQ ID No. 10.

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variant

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Loop 8 modifications

In a further embodiment the invention relates to a variant of a parent Termamyl-like α-amylase, in which variant at least one of the amino acid residues of the parent α-amylase, which is present in a fragment corresponding to amino acid residues 322-346 of the amino acid sequence of SEQ ID No. 2, has/have been deleted or replaced with one or more amino acid residues, which is/are present in an amino acid fragment corresponding to amino acid residues 291-313 of the amino acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has/have been inserted using the relevant part of SEQ ID No. 10 or a corresponding part of another Fungamyl-like α-amylase as a template.

- 25 For instance, the variant may be one, in which the amino acid fragment X-Y of the parent α -amylase, which corresponds to or is within the amino acid fragment 322-346 of SEQ ID No. 2, has/have been replaced with an amino acid fragment Z-V, which corresponds to or is within the amino acid fragment 291-313 of the amino acid sequence shown in SEQ ID No. 10, in which
 - X is an amino acid residue corresponding to the amino acid occupying position 322, 323, 324 or 325 of SEQ ID No. 2,

Y is an amino acid residue corresponding to the amino acid occupying position 343, 344, 345 or 346 of SEQ ID No. 2,

Z is an amino acid residue corresponding to the amino acid occupying position 291, 292, 293 or 294 of SEQ ID No. 10, and

V is an amino acid residue corresponding to the amino acid occupying position 310, 311, 312 or 313 of SEQ ID No. 10.

A specific variant according to this aspect of the invention is one, in which the amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 325-345 of SEQ D No. 10 2, has been replaced with the amino acid fragment corresponding to amino acid residues 294-313 of the amino acid sequence shown in SEQ ID No. 10.

Ca2+ dependency

15 It is highly desirable to be able to decrease the Ca² dependency of a Termamyl-like α-amylase. Accordingly, in a further aspect the invention relates to a variant of a parent Termamyl-like α-amylase, which exhibits α-amylase activity and which has a decreased Ca² dependency as compared to the parent α-amylase. The decreased Ca² dependency has the functional result that the variant exhibits a satisfactory amylolytic activity in the presence of a lower concentration of calcium ion in the extraneous medium than is necessary for the parent enzyme and, for example, therefore is less sensitive than the 25 parent to calcium ion-depleting conditions such as those obtained in media containing calcium-complexing agents (such as certain detergent builders).

The decreased Ca^{2+} dependency of the variant of the invention 10 may advantageously be achieved by increasing the Ca^{2+} binding affinity of the parent Termamyl-like α -amylase, in other words the stronger the Ca^{2+} binding of the enzyme, the lower is the Ca^{2+} dependency.

35 It is presently believed that amino acid residues located within 10Å from a sodium or calcium ion are involved in or are of importance for the Ca² binding capability of the enzyme.

Accordingly, the variant according to this aspect of the invention is preferably one, which has been modified in one or more amino acid residues present within 10Å from a calcium and/or sodium in identified in the three-dimensional Termamylika resultant actions in such a manager that the efficiency of such a such as the efficiency of such as the efficiency of

5 like α -amylase structure in such a manner that the affinity of the α -amylase for calcium is increased.

The amino acid residues found within a distance of 10Å from the Ca^{2^*} binding sites of the B. licheniformis α -amylase with the 10 amino acid sequence SEQ ID NO 2 were determined as described in Example 2 and are as follows:

V102, I103, N104, H105, K106, R125, W155, W157, Y158, H159, F160, D161, G162, T163, Y175, K176, F177, G178, K180, A181, 19 W182, D183, W184, E185, V186, S187, N192, Y193, D194, Y195, L196, M197, Y198, A199, D200, I201, D202, Y203, D204, H205, P206, V208, A209, D231, A232, V233, K234, H235, I236, K237, F238, F240, L241, A294, A295, S296, T297, Q298, G299, G300, G301, Y302, D303, M304, R305, K306, L307, W342, F343, L346, G415, D416, S417, V419, A420, N421, S422, G423, L424, I428, T429, D430, G431, P432, V440, G441, R442, Q443, N444, A445, G446, E447, T448, W449, I462, G475, Y480, V481, Q482, R483.

- 25 In order to construct a variant according to this aspect of the invention it is desirable to replace at least one of the above mentioned amino acid residues (or an amino acid residue occupying an equivalent position in another Termamyl-like α-amylase than that defined by SEQ ID NO 2), which is contemplated to be involved in providing a non-optimal calcium binding, with any other amino acid residue which improves the Ca² binding affinity of the variant enzyme. In practice, the identification and subsequent modification of the amino acid residue is performed by the following method:
 - i) identifying an amino acid residue within 10Å from a Ca^2 -binding site of a Termamyl-like α -amylase structure, which from

structural or functional considerations is believed to be responsible for a non-optimal calcium ion interaction,

- ii) constructing a variant in which said amino acid residue is 5 replaced with another amino acid residue which from structural or functional considerations is believed to be important for establishing a higher Ca^{2*} binding affinity, and testing the Ca^{2*} dependency of the resulting Termamyl-like a-amylase variant.
- 10 In the present context, the term "non-optimal calcium ion interaction" is intended to indicate that the amino acid residue in question is selected on the basis of a presumption that substituting said amino acid residue for another may improve a calcium ion binding interaction of the enzyme. For instance, the amino acid residue in question may be selected on the basis of one or more of the following considerations:
- to obtain an improved interaction between a calcium ion and an amino acid residue located near to the surface of the enzyme (as identified from the structure of the Termamyl-like α -amylase). For instance, if the amino acid residue in question is exposed to a surrounding solvent, it may be advantageous to increase the shielding of said amino acid residue from the solvent so as to provide for a stronger interaction between 25 said amino acid residue and a calcium ion. This can be achieved by replacing said residue (or an amino acid residue in the vicinity of said residue contributing to the shielding) by an amino acid residue which is more bulky or otherwise results in an improved shielding effect.

- to stabilize a calcium binding site, for instance by stabilizing the structure of the Termamyl-like α -amylase (e.g. by stabilizing the contacts between the A, B and C domains or stabilizing one or more of the domains as such). This may,

35 e.g., be achieved by providing for a better coordination to amino acid side chains, which may, e.g., be obtained by replacing an N residue with a D residue and/or a Q residue with

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an E residue (e.g. N104D), e.g. within 10Å, and preferably within 3 or 4Å, of a calcium binding site.

- to protect the calcium binding site or to improve the s coordination between the calcium ion and the calcium binding site, e.g. by providing a stronger interaction between the ion and the binding site.

Before actually constructing a Termamyl-like α -amylase variant 10 according to the above principles it may be convenient to evaluate the contemplated amino acid modification by its accommodation into the Termamyl-like α -amylase structure, e.g. into a model structure of the parent Termamyl-like α -amylase.

15 Preferably, the amino acid residue to be modified is located within 8Å of a Ca²⁺ binding site residue, such as within 5Å of such residue. The amino acid residues within 8Å and 5Å, respectively, may easily be identified by an analogous method used for identifying amino acid residues within 10Å (cf. 20 Example 2).

The following mutation is contemplated to be of particular interest with respect to decreasing the Ca^{2*} dependency of a Termamyl-like α -amylase:

25 N104D (of the B. licheniformis α -amylase SEQ ID NO 2, or an equivalent (N to D) mutation of an equivalent position in another Termamyl-like α -amylase.)

In connection with substitutions of relevance for Ca² dependency, some other substitutions appear to be of importance in stabilizing the enzyme conformation (for instance the Domains A-B and/or Domains A-C interactions contributing to the overall stability of the enzyme) in that they may, e.g., enhance the strength of binding or retention of calcium ion or sodium ion at or within a calcium or sodium binding site, respectively, within the parent Termamyl-like α-amylase.

It is desirable to stabilize the C-domain in order to increase the calcium stability and/or thermostability of the enzyme. In this connection the stabilization may result in a stabilization of the binding of calcium by the enzyme, and an improved 5 contact between the C-domain and the A-domain (of importance for thermostability). The latter may be achieved by introduction of cystein bridges, salt bridges or increase

10 For instance, the C-domain of the B. licheniformis α-amylase having the amino acid sequence shown in SEQ ID No. 2 may be stabilized by introduction of a cystein bridge between domain A and domain C, e.g. by introducing of the following mutations: A349C+I479C and/or L346C+I430C.

hydrogen, hydrophobic and/or electrostatic interactions.

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A salt bridge may be obtained by introduction of the following mutations:

N457D.E

N457D, E+K385R

20 F350D,E+I430R,K
F350D,E+I411R,K

The calcium site of Domain C may be stabilized by replacing the amino acid residues H408 and/or G303 with any other amino acid 25 residue. Of particular interest is the following mutations:

H408Q,E,N,D and/or G303N,D,Q,E

which are contemplated to provide a better calcium binding or protection from calcium depletion.

30 Similar mutations may be introduced in equivalent positions of other Termamyl-like α -amylases.

Other substitution mutations (relative to B. licheniformis α -amylase, SEQ ID No. 2) which appear to be of importance,

35 inter alia, in the context of reducing calcium dependency include the following: R23K, H156Y, A181T, A209V and G310D (or equivalent mutations in equivalent positions in another Termamyl-like α -amylase). Substitutions of R214 and P345 with

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other amino acids may also be of importancen in this connection.

Variants with altered activity at higher/lower pH

- It is contemplated that it is possible to change the pH optima of a Termamyl-like α -amylase or the enzymatic activity at a given pH by changing the pKa of the active site residues. This may be achieved, e.g. by changing the electrostatic interaction or hydrophobic interaction between functional groups of amino acid side chains of the amino acid residue to be modified and of its close surroundings. This may, e.g., be accomplished by the following method:
- 15 i) in a structure of the Termamyl-like α -amylase in question to identifying an amino acid residue within 15Å from an active site residue, in particulular 10Å from an active site residue, which amino acid residue is contemplated to be involved in electrostatic or hydrophobic interactions with an active site residue.
- ii) replacing, in the structure, said amino acid residue with an amino acid residue which changes the electrostatic and/or hydrophobic surroundings of an active site residue and sevaluating the accommodation of the amino acid residue in the structure,
- iii) optionally repeating step i) and/or ii) until an amino acid replacement has been identified which is accommodated into the structure,
 - iv) constructing a Termamyl-like α -amylase variant resulting from steps i), ii) and optionally iii) and testing the pH dependent enzymatic activity of interest of said variant.
 - In the above method it may be of particular relevance to add a positively charged residue within 5Å of a glutamate (thereby lowering the pKa of the glutamate from about 4.5 to 4), or to

add a negatively charged residue within 5 Å of a glutamate (thereby increasing the pKa to about 5), or to make similar modifications within a distance of about 5\AA of a Histidine.

5 In a further aspect the invention relates to a variant of a Termamyl-like α -amylase which exhibits a higher activity at a lower pH (e.g. compared to the pH optimum) than the parent α -amylase. In particular, the variant comprises a mutation of an amino acid residue corresponding to at least one of the following positions of the B. licheniformis α -amylase (SEQ ID NO 2):

E336, Q333, P331, I236, V102, A232, I103, L196

The following mutations are of particular interest:

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E336R,K Q333R,K P331R,K V102R,K,A,T,S,G;

20 I236K,R,N; I103K,R; L196K,R; A232T.S.G:

25 or any combination of two or more of these variants or any combination of one or more of these variants with any of the other variants disclosed herein.

In a still further aspect the invention relates to a variant of a Termamyl-like α -amylase which has a higher activity at a higher pH than the parent α -amylase. In particular, the variant comprises a mutation of an amino acid residue corresponding to at least one of the following positions of the B. licheniformis α -amylase (SEQ ID NO 2):

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N236, H281, Y273

In particular, the variant comprises a mutation corresponding to at least one of the following mutations of the B. licheniformis α -amylase (SEQ ID NO 2):

5 N326I,Y,F,L,V H281F,I,L Y273F,W

or any combination of two or more of these variants or any combination of one or more of these variants with any of the other variants disclosed herein.

A mutation which appears to be importance in relation to the specific activity of variants of the invention is a mutation is corresponding to the substitution S187D in B. licheniformis α -amylase (SEQ ID NO 2).

Variants with increased thermostability and/or altered temperature optimum

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In a further desired aspect the invention relates to a variant of a parent Termamyl-like α -amylase, which variant is the result of one or more amino acid residues having been deleted from, replaced or added to the parent α -amylase so as to obtain an increased thermostability of the variant.

The Termamyl-like α -amylase structure contains a number of unique internal holes, which may contain water, and a number of crevices. In order to increase the thermostability of the α -30 amylase it may be desirable to reduce the number of holes and crevices (or reduce the size of the holes or crevices), e.g. by introducing one or more hydrophobic contacts, preferably achieved by introducing bulkier residues, in the vicinity or surroundings of the hole. For instance, the amino acid residues to be modified are those which are involved in the formation of the hole.

Accordingly, in a further aspect the present invention relates to a method of increasing the thermostability and/or altering the temperature optimum of a parent Termamyl-like α -amylase, which method comprises

- i) identifying an internal hole or a crevice of the parent Termamyl-like α -amylase in the three-dimensional structure of said α -amylase,
- 10 ii) replacing, in the structure, one or more amino acid residues in the neighbourhood of the hole or crevice identified in i) with another amino acid residue which from structural or functional considerations is believed to increase the hydrophobic interaction and to fill out or reduce the size of 15 the hole or crevice.
 - iii) constructing a Termamyl-like α -amylase variant resulting from step ii) and testing the thermostability and/or temperature optimum of the variant.
- 20 The structure used for identifying the hole or crevice of the parent Termamyl-like α -amylase may be the structure identified in Appendix 1 or a model structure of the parent Termamyl-like α -amylase built thereon.
- 25 It will be understood that the hole or crevice is identified by the amino acid residues surrounding the hole/crevice, and that modification of said amino acid residues are of importance for filling or reducing the size of the hole/crevice. The particular amino acid residues referred to below are those 30 which in crystal structure have been found to flank the hole/crevice in question.
- In order to fill (completely or partly) a major hole located between domain A and B, mutation to any other amino acid 35 residue of an amino acid residue corresponding to one or more of the following residues of the B. licheniformis α -amylase (SEQ ID NO 2) is contemplated:

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L61, Y62, F67, K106, G145, I212, S151, R214, Y150, F143, R146

Of particular interest is a mutation to a more bulky amino acid residue than the amino acid residue of the parent enzyme.

Of particular interest is a variant of a Termamyl-like α -10 amylase which comprises a mutation corresponding to the following mutations (using the numbering of B. licheniformis α -amylase (SEQ ID NO 2):

L61W, V, F;

15 Y62W;

F67W:

K106R, F, W;

G145F.W

I212F, L, W, Y, R, K;

20 S151 replaced with any other amino acid residue and in particular with F,W,I or L; R214W;

Y150R,K;

F143W; and/or

25 R146W.

In order to fill a hole in the vicinity of the active site mutation to any other amino acid residue of an amino acid residue corresponding to one or more of the following residues of the B. licheniformis α -amylase (SEQ ID NO 2) is contemplated:

L241. I236.

35 Of interest is a mutation to a more bulky amino acid residue.

Of particular interest is a variant of a Termamyl-like α -amylase which comprises a mutation corresponding to one or more of the following mutations in the B. licheniformis α -amylase:

5 L241I,F,Y,W; and/or I236L,F,W,Y

In order to fill a hole in the vicinity of the active site mutation to any other amino acid residue of an amino acid residue corresponding to one or more of the following residues of the B. licheniformis α -amylase (SEQ ID NO 2) is contemplated: L7, V259, F284

15 Of interest is a mutation to a more bulky amino acid residue.

Of particular interest is a variant of a Termamyl-like α -amylase which comprises a mutation corresponding to one or more of the following mutations in the B. licheniformis α -amylase:

L7F,I,W V259F,I,L F284W

25 In order to fill a hole in the vicinity of the active site mutation to any other amino acid residue of an amino acid residue corresponding to one or more of the following residues of the B. licheniformis α -amylase (SEQ ID NO 2) is contemplated:

30 F350, F343

Of interest is a mutation to a more bulky amino acid residue.

35 Of particular interest is a variant of a Termamyl-like α -amylase which comprises a mutation corresponding to one or more of the following mutations in the B. licheniformis α -amylase: F350W

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F343W

In order to fill a hole in the vicinity of the active site mutation to any other amino acid residue of an amino acid s residue corresponding to one or more of the following residues of the B. licheniformis α -amylase (SEQ ID NO 2) is contemplated:

L427, V481

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Of interest is a mutation to a more bulky amino acid residue.

Of particular interest is a variant of a Termamyl-like α -amylase which comprises a mutation corresponding to one or more 15 of the following mutations in the B. licheniformis α -amylase:

L427F, L, W V481, F, I, L, W

20 Variants with an altered cleavage pattern

In the starch liquefaction process it is desirable to use an α-amylase which is capable of degrading the starch molecules into long branched oligo saccharides (like, e.g. the Fungamyl-like α-amylases) rather than shorter branched oligo saccharides (like conventional Termamyl-like α-amylases). The resulting very small branched oligosaccharides (panose precursors) cannot be hydrolyzed properly by pullulanases, which in the liquefaction process are used after the α-amylases and before the amyloglucosidases. Thus, in the presence of panose precursors the action of amylo-glucoamylase ends up with a high degree of the small branched limiting-dextrin, the trisaccharide panose. The presence of panose lowers the saccharification yield significantly and is thus undesirable.

35 Thus, one aim of the present invention is to change the degradation characteristics of a Termamyl-like α -amylase to that of a Fungamyl-like α -amylases without at the same time reducing the thermostability of the Termamyl-like α -amylase.

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Accordingly, in a further aspect the invention relates to a variant of a Termamyl-like α -amylase which has a reduced ability to cleave a substrate close to the branching point.

- 5 The variant may suitably be constructed by a method which comprises
- i) identifying the substrate binding area of the parent Termamyl-like α -amylase in a model of the three-dimensional structure of said α -amylase, (e.g. within a sphere of 4Å from the substrate binding site (as defined in the section above entitled "Substrate Binding Site"),
- ii) replacing, in the model, one or more amino acid residues of
 15 the substrate binding area of the cleft identified in i), which is/are believed to be responsible for the cleavage pattern of the parent α-amylase, with another amino acid residue which from structural considerations is believed to result in an altered substrate cleavage pattern, or deleting one or more
 20 amino acid residues of the substrate binding area contemplated to introduce favourable interactions to the substrate or adding one or more amino acid residues to the substrate binding area contemplated to introduce favourable interactions to the substrate, and
- 25 iii) constructing a Termamyl-like α -amylase variant resulting from step ii) and testing the substrate cleavage pattern of the variant.
- Of particular interest is a variant which cleaves an 30 amylopectin substrate, from the reducing end, more than one glucose unit from the branching point, preferably more than two or three glucose units from the branching point, i.e. at a further distance from the branching point than that obtained by use of a wild type B. licheniformis α-amylase.

Residues of particular interest in connection with this aspect of the invention correspond to the following residues of the B. licheniformis α -amylase (SEQ ID NO 2): V54, D53, Y56, Q333,

G57, and the variants according to this aspect preferably comprises a mutation in one or more of these residues.

In particular, the variant comprises at least one of the 5 following mutations, which are expected to prevent cleavage close to the branching point:

V54L,I,F,Y,W,R,K,H,E,Q
D53L,I,F,Y,W

10 Y56W
Q333W
G57all possible amino acid residues
A52amino acid residues larger than A, e.g. A52W,Y,L,F,I.

15 Variants of a fungal α-amylase

In a still further embodiment the invention relates to a variant of a parent Fungamyl-like α -amylase, in which variant at least one of the amino acid residues of the parent α -20 amylase, which is/are present in an amino acid fragment corresponding to amino acid residues 291-313 of the amino acid sequence of SEQ ID No. 10, has/have been deleted or replaced with one or more of the amino acid residues, which is/are present in an amino acid fragment corresponding to amino acid residues 98-210 of the amino acid sequence shown in SEQ ID No. 4, or in which one or more additional amino acid residues has/have been inserted using the relevant part of SEQ ID No. 4 or a corresponding part of another Termamyl-like α -amylase as a template.

For instance, the variant may be one, in which the amino acid fragment X-Y of the parent α-amylase, which corresponds to or is within the amino acid fragment 117-185 of SEQ ID No. 10, has/have been replaced with an amino acid fragment Z-V, which corresponds to or is within the amino acid fragment 98-210 of the amino acid sequence shown in SEQ ID No. 4, in which variant

- X is an amino acid residue corresponding to the amino acid occupying position 117, 118, 119, 120 or 121 of SEQ ID No. 10,
- Y is an amino acid residue corresponding to the amino acid cocupying position 181, 182, 183, 184 or 185 of SEQ ID No. 10,
 - Z is an amino acid residue corresponding to the amino acid occupying position 98, 99, 100, 101 or 102 of SEQ ID No. 4, and
- 10 V is an amino acid residue corresponding to the amino acid occupying position 206, 207, 208, 209 or 210 of SEQ ID No. 4.
- A specific example of a variant according to this aspect of the invention is one, in which the amino acid fragment of the 15 parent α-amylase, which corresponds to amino acid residues 121-181 of SEQ ID No. 10, has been replaced with the amino acid fragment corresponding to amino acid residues 102-206 of the amino acid sequence shown in SEQ ID No. 4.
- 20 Another example of a variant according to this aspect of the invention is one, in which the amino acid fragment of the parent α-amylase, which corresponds to amino acid residues 121-174 of SEQ ID No. 10, has been replaced with the amino acid fragment corresponding to amino acid residues 102-199 of the 25 amino acid sequence shown in SEQ ID No. 4.
- In a further embodiment the invention relates to a variant of a parent Fungamyl-like α -amylase, in which an amino acid fragment corresponding to amino acid residues 181-184 of the amino acid sequence shown in SEQ ID No. 10 has been deleted.

General mutations in variants of the invention

It may be preferred that the variant of the invention or prepared in accordance with the method of the invention comprises one or more modifications in addition to those outlined above. Thus, it may be advantageous that one or more proline residues present in the part of the α -amylase variant

having been modified is/are replaced with a non-proline residue which may be any of the possible, naturally occurring non-proline residues, and which preferably is an alanine, glycine, serine, threonine, valine or leucine.

Analogously, it may be preferred that one or more cysteine residues present in the amino acid residues with which the parent α -amylase is modified are replaced with a non-cysteine residues such as serine, alanine, threonine, glycine, valine or leucine.

Furthermore, the variant of the invention may either as the only modification or in combination with any of the above outlined modifications be modified so that one or more Asp 15 and/or Glu present in an amino acid fragment corresponding to the amino acid fragment 185-209 of SEQ ID No. 8 is replaced by an Asn and/or Gln, respectively. Also of interest is the modification of one or more of the Lys residues present in the Termamyl-like α-amylase is replaced by an Arg present in an 20 amino acid fragment corresponding to the amino acid fragment 185-209 of SEQ ID No. 8 is replaced by an Asn and/or Gln, respectively.

It will be understood that in accordance with the present invention variants may be prepared which carry two or more of the above outlined modifications. For instance, variants may be prepared which comprises a modification in the loop 1 and loop 2 region, a modification in loop 2 and limited loop 3, a modification in loop 1, loop 3 and loop 8, etc.

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Furthermore, it may be advantageous to introduce pointmutations in any of the variants described herein.

Methods of preparing α-amylase variants

35 Several methods for introducing mutations into genes are known in the art. After a brief discussion of the cloning of αamylase-encoding DNA sequences, methods for generating

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mutations at specific sites within the $\alpha\text{-amylase-encoding}$ sequence will be discussed.

Cloning a DNA sequence encoding an \u03c3-amylase

- 5 The DNA sequence encoding a parent α -amylase may be isolated from any cell or microorganism producing the α -amylase in question, using various methods well known in the art. First, a genomic DNA and/or cDNA library should be constructed using chromosomal DNA or messenger RNA from the organism that produces the α -amylase to be studied. Then, if the amino acid sequence of the α -amylase is known, homologous, labelled oligonucleotide probes may be synthesized and used to identify α -amylase-encoding clones from a genomic library prepared from the organism in question. Alternatively, a labelled oligonulaction probe containing sequences homologous to a known α -amylase gene could be used as a probe to identify α -amylase-encoding clones, using hybridization and washing conditions of lower stringency.
- Yet another method for identifying α -amylase-encoding clones would involve inserting fragments of genomic DNA into an expression vector, such as a plasmid, transforming α -amylase-negative bacteria with the resulting genomic DNA library, and then plating the transformed bacteria onto agar containing a substrate for α -amylase, thereby allowing clones expressing the α -amylase to be identified.

Alternatively, the DNA sequence encoding the enzyme may be prepared synthetically by established standard methods, e.g. 30 the phosphoroamidite method described by S.L. Beaucage and M.H. Caruthers (1981) or the method described by Matthes et al. (1984). In the phosphoroamidite method, oligonucleotides are synthesized, e.g. in an automatic DNA synthesizer, purified, annealed, ligated and cloned in appropriate vectors.

Finally, the DNA sequence may be of mixed genomic and synthetic origin, mixed synthetic and cDNA origin or mixed genomic and cDNA origin, prepared by ligating fragments of synthetic,

genomic or cDNA origin (as appropriate, the fragments corresponding to various parts of the entire DNA sequence), in accordance with standard techniques. The DNA sequence may also be prepared by polymerase chain reaction (PCR) using specific 5 primers, for instance as described in US 4,683,202 or R.K. Saiki et al. (1988).

Site-directed mutagenesis

Once an α -amylase-encoding DNA sequence has been isolated, and 10 desirable sites for mutation identified, mutations may be introduced using synthetic oligonucleotides. These oligonucleotides contain nucleotide sequences flanking the desired mutation sites; mutant nucleotides are inserted during oligonucleotide synthesis. In a specific method, a single-stranded 15 gap of DNA, bridging the α-amylase-encoding sequence, is created in a vector carrying the \alpha-amylase gene. Then the synthetic nucleotide, bearing the desired mutation, is annealed to a homologous portion of the single-stranded DNA. The remaining gap is then filled in with DNA polymerase I (Klenow fragment) 20 and the construct is ligated using T4 ligase. A specific example of this method is described in Morinaga et al. (1984). US 4,760,025 discloses the introduction of oligonucleotides encoding multiple mutations by performing minor alterations of the cassette. However, an even greater variety of mutations can 25 be introduced at any one time by the Morinaga method, because a multitude of oligonucleotides, of various lengths, can be introduced.

Another method of introducing mutations into α -amylase-encoding DNA sequences is described in Nelson and Long (1989). It involves the 3-step generation of a PCR fragment containing the desired mutation introduced by using a chemically synthesized DNA strand as one of the primers in the PCR reactions. From the PCR-generated fragment, a DNA fragment carrying the mutation $_{35}$ may be isolated by cleavage with restriction endonucleases and reinserted into an expression plasmid.

Random mutagenesis

Random mutagenesis is suitably performed either as localized or region-specific random mutagenesis in at least three parts of the gene translating to the amino acid sequence shown in question, or within the whole gene.

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For region-specific random mutagenesis with a view to improving the thermal stability of a parent Termamyl-like α -amylase, codon positions corresponding to the following amino acid residues of the *B. licheniformis* α -amylase (SEQ ID NO 2) may 10 appropriately be targeted:

To improve the stability of the calcium site between Domain A and C

I428-A435

15 T297-L308

F403-V409

To improve the stability between domain A and B: D180-D204

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20 H156-T163

A232-F238

With a view to achieving improved binding of a substrate (i.e. improved binding of a carbohydrate species, such as amylose or amylopectin) by a Termamyl-like α -amylase variant, modified (e.g. higher) substrate specificity and/or modified (e.g. higher) specificity with respect to cleavage (hydrolysis) of substrate, it appears that the following codon positions for the amino acid sequence shown in SEQ ID NO 2 (or equivalent codon positions for another parent Termamyl-like α -amylase in the context of the invention) may particularly appropriately be targeted:

13-18

35 50-56

70-76

102-109

163-172

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189-199 229-235 360-264 327-335

The random mutagenesis of a DNA sequence encoding a parent α amvlase to be performed in accordance with step a) of the above-described method of the invention may conveniently be performed by use of any method known in the art.

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For instance, the random mutagenesis may be performed by use of a suitable physical or chemical mutagenizing agent, by use of a suitable oligonucleotide, or by subjecting the DNA sequence to PCR generated mutagenesis. Furthermore, the random mutagene-15 sis may be performed by use of any combination of these mutagenizing agents.

The mutagenizing agent may, e.g., be one which induces transitions, transversions, inversions, scrambling, deletions, 20 and/or insertions.

Examples of a physical or chemical mutagenizing agent suitable for the present purpose include ultraviolet (UV) irradiation, hydroxylamine, N-methyl-N'-nitro-N-nitrosoguanidine (MNNG), O-25 methyl hydroxylamine, nitrous acid, ethyl methane sulphonate (EMS), sodium bisulphite, formic acid, and nucleotide analoques.

When such agents are used, the mutagenesis is typically per-30 formed by incubating the DNA sequence encoding the parent enzyme to be mutagenized in the presence of the mutagenizing agent of choice under suitable conditions for the mutagenesis to take place, and selecting for mutated DNA having the desired properties.

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When the mutagenesis is performed by the use of an oligonucleotide, the oligonucleotide may be doped or spiked with the three non-parent nucleotides during the synthesis of the oligonucleo-

tide at the positions which are to be changed. The doping or spiking may be done so that codons for unwanted amino acids are avoided. The doped or spiked oligonucleotide can be incorporated into the DNA encoding the amylolytic enzyme by any 5 published technique, using e.g. PCR, LCR or any DNA polymerase and ligase.

When PCR-generated mutagenesis is used, either a chemically treated or non-treated gene encoding a parent α -amylase enzyme 10 is subjected to PCR under conditions that increase the misincorporation of nucleotides (Deshler 1992; Leung et al., Technique, Vol.1, 1989, pp. 11-15).

A mutator strain of *E. coli* (Fowler et al., Molec. Gen. Genet., 133, 1974, pp. 179-191), *S. cereviseae* or any other microbial organism may be used for the random mutagenesis of the DNA encoding the amylolytic enzyme by e.g. transforming a plasmid containing the parent enzyme into the mutator strain, growing the mutator strain with the plasmid and isolating the mutated plasmid from the mutator strain. The mutated plasmid may subsequently be transformed into the expression organism.

The DNA sequence to be mutagenized may conveniently be present in a genomic or cDNA library prepared from an organism expressing the parent amylolytic enzyme. Alternatively, the DNA sequence may be present on a suitable vector such as a plasmid or a bacteriophage, which as such may be incubated with or otherwise exposed to the mutagenizing agent. The DNA to be mutagenized may also be present in a host cell either by being integrated in the genome of said cell or by being present on a vector harboured in the cell. Finally, the DNA to be mutagenized may be in isolated form. It will be understood that the DNA sequence to be subjected to random mutagenesis is preferably a cDNA or a genomic DNA sequence.

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In some cases it may be convenient to amplify the mutated DNA sequence prior to the expression step (b) or the screening step (c) being performed. Such amplification may be performed in

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accordance with methods known in the art, the presently preferred method being PCR-generated amplification using oligonucleotide primers prepared on the basis of the DNA or amino acid sequence of the parent enzyme.

Subsequent to the incubation with or exposure to the mutagenizing agent, the mutated DNA is expressed by culturing a
suitable host cell carrying the DNA sequence under conditions
allowing expression to take place. The host cell used for this
purpose may be one which has been transformed with the mutated
DNA sequence, optionally present on a vector, or one which was
carried the DNA sequence encoding the parent enzyme during the
mutagenesis treatment. Examples of suitable host cells are the
following: grampositive bacteria such as Bacillus subtilis,
Bacillus licheniformis, Bacillus lentus, Bacillus brevis,
Bacillus stearothermophilus, Bacillus alkalophilus, Bacillus
amyloliquefaciens, Bacillus coagulans, Bacillus circulans,
Bacillus lautus, Bacillus megaterium, Bacillus thuringiensis,
Streptomyces lividans or Streptomyces murinus; and gramnegative
bacteria such as E.coli.

The mutated DNA sequence may further comprise a DNA sequence encoding functions permitting expression of the mutated DNA sequence.

Localized random mutagenesis: the random mutagenesis may advantageously be localized to a part of the parent α-amylase in question. This may, e.g., be advantageous when certain regions of the enzyme have been identified to be of particular modified are expected to result in a variant having improved properties. Such regions may normally be identified when the tertiary structure of the parent enzyme has been elucidated and related to the function of the enzyme.

The localized random mutagenesis is conveniently performed by use of PCR- generated mutagenesis techniques as described above or any other suitable technique known in the art.

Alternatively, the DNA sequence encoding the part of the DNA sequence to be modified may be isolated, e.g. by being inserted into a suitable vector, and said part may subsequently be subjected to mutagenesis by use of any of the mutagenesis methods discussed above.

With respect to the screening step in the above-mentioned method of the invention, this may conveniently performed by use of aa filter assay based on the following principle:

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A microorganism capable of expressing the mutated amylolytic enzyme of interest is incubated on a suitable medium and under suitable conditions for the enzyme to be secreted, the medium being provided with a double filter comprising a first protein15 binding filter and on top of that a second filter exhibiting a low protein binding capability. The microorganism is located on the second filter. Subsequent to the incubation, the first filter comprising enzymes secreted from the microorganisms is separated from the second filter comprising the microorganisms.

20 The first filter is subjected to screening for the desired enzymatic activity and the corresponding microbial colonies present on the second filter are identified.

The filter used for binding the enzymatic activity may be any 25 protein binding filter e.g. nylon or nitrocellulose. The top-filter carrying the colonies of the expression organism may be any filter that has no or low affinity for binding proteins e.g. cellulose acetate or DuraporeTM. The filter may be pretreated with any of the conditions to be used for screening or may be treated during the detection of enzymatic activity.

The enzymatic activity may be detected by a dye, fluorescence, precipitation, pH indicator, IR-absorbance or any other known technique for detection of enzymatic activity.

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The detecting compound may be immobilized by any immobilizing agent e.g. agarose, agar, gelatine, polyacrylamide, starch, filter paper, cloth; or any combination of immobilizing agents.

 α -Amylase activity is detected by Cibacron Red labelled amylopectin, which is immobilized on agarose. For screening for variants with increased thermal and high-pH stability, the filter with bound α -amylase variants is incubated in a buffer 5 at pH 10.5 and 60° or 65°C for a specified time, rinsed briefly in deionized water and placed on the amylopectin-agarose matrix for activity detection. Residual activity is seen as lysis of Cibacron Red by amylopectin degradation. The conditions are chosen to be such that activity due to the α -amylase having the 10 amino acid sequence shown in SEQ ID No.1 can barely be detected. Stabilized variants show, under the same conditions, increased colour intensity due to increased liberation of Cibacron Red

15 For screening for variants with an activity optimum at a lower temperature and/or over a broader temperature range, the filter with bound variants is placed directly on the amylopectin-Cibacron Red substrate plate and incubated at the desired temperature (e.g. 4°C, 10°C or 30°C) for a specified time.
20 After this time activity due to the \(\alpha\)-amylase having the amino acid sequence shown in SEQ ID No.1 can barely be detected, whereas variants with optimum activity at a lower temperature will show increase amylopectin lysis. Prior to incubation onto the amylopectin matrix, incubation in all kinds of desired 25 media - e.g. solutions containing Ca^{2*}, detergents, EDTA or other relevant additives - can be carried out in order to screen for changed dependency or for reaction of the variants in question with such additives.

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Testing of variants of the invention

The testing of variants of the invention may suitably be performed by determining the starch-degrading activity of the 35 variant, for instance by growing host cells transformed with a DNA sequence encoding a variant on a starch-containing agarose plate and identifying starch-degrading host cells. Further testing as to altered properties (including specific activity,

substrate specificity, cleavage pattern, thermoactivation, pH optimum, pH dependency, temperature optimum, and any other parameter) may be performed in accordance with methods known in the art.

Expression of \alpha-amylase variants

According to the invention, a DNA sequence encoding the variant produced by methods described above, or by any alternative methods known in the art, can be expressed, in enzyme form, 10 using an expression vector which typically includes control sequences encoding a promoter, operator, ribosome binding site, translation initiation signal, and, optionally, a repressor gene or various activator genes.

15 The recombinant expression vector carrying the DNA sequence encoding an α-amylase variant of the invention may be any vector which may conveniently be subjected to recombinant DNA procedures, and the choice of vector will often depend on the host cell into which it is to be introduced. Thus, the vector may be an autonomously replicating vector, i.e. a vector which exists as an extrachromosomal entity, the replication of which is independent of chromosomal replication, e.g. a plasmid, a bacteriophage or an extrachromosomal element, minichromosome or an artificial chromosome. Alternatively, the vector may be one which, when introduced into a host cell, is integrated into the host cell genome and replicated together with the chromosome(s) into which it has been integrated.

In the vector, the DNA sequence should be operably connected to a suitable promoter sequence. The promoter may be any DNA sequence which shows transcriptional activity in the host cell of choice and may be derived from genes encoding proteins either homologous or heterologous to the host cell. Examples of suitable promoters for directing the transcription of the DNA sequence encoding an α -amylase variant of the invention, especially in a bacterial host, are the promoter of the lac operon of E.coli, the Streptomyces coelicolor agarase gene daga promoters, the promoters of the Bacillus licheniformis α -

amylase gene (amyL), the promoters of the Bacillus stearothermophilus maltogenic amylase gene (amyM), the promoters of the Bacillus amyloliquefaciens α -amylase (amyQ), the promoters of the Bacillus subtilis xylA and xylB genes etc. For transcription in a fungal host, examples of useful promoters are those derived from the gene encoding A. oryzae TAKA amylase, Rhizomucor miehei aspartic proteinase, A. niger neutral α -amylase, A. niger acid stable α -amylase, A. niger glucoamylase, Rhizomucor miehei lipase, A. oryzae alkaline protease, A. oryzae triose phosphate isomerase or A. nidulans acetamidase.

The expression vector of the invention may also comprise a suitable transcription terminator and, in eukaryotes, poly- is adenylation sequences operably connected to the DNA sequence encoding the α -amylase variant of the invention. Termination and polyadenylation sequences may suitably be derived from the same sources as the promoter.

- 20 The vector may further comprise a DNA sequence enabling the vector to replicate in the host cell in question. Examples of such sequences are the origins of replication of plasmids pUC19, pACYC177, pUB110, pE194, pAMB1 and pIJ702.
- 25 The vector may also comprise a selectable marker, e.g. a gene the product of which complements a defect in the host cell, such as the dal genes from B. subtilis or B. licheniformis, or one which confers antibiotic resistance such as ampicillin, kanamycin, chloramphenicol or tetracyclin resistance. Fursion thermore, the vector may comprise Aspergillus selection markers such as amdS, argB, niaD and sC, a marker giving rise to hygromycin resistance, or the selection may be accomplished by co-transformation, e.g. as described in WO 91/17243.
- 35 While intracellular expression may be advantageous in some respects, e.g. when using certain bacteria as host cells, it is generally preferred that the expression is extracellular. In general, the Bacillus α -amylases mentioned herein comprise a

preregion permitting secretion of the expressed protease into the culture medium. If desirable, this preregion may be replaced by a different preregion or signal sequence, conveniently accomplished by substitution of the DNA sequences encoding the respective preregions.

The procedures used to ligate the DNA construct of the invention encoding an α -amylase variant, the promoter, terminator and other elements, respectively, and to insert them into suitable vectors containing the information necessary for replication, are well known to persons skilled in the art (cf., for instance, Sambrook et al. (1989)).

The cell of the invention, either comprising a DNA construct or 15 an expression vector of the invention as defined above, is advantageously used as a host cell in the recombinant production of an α-amylase variant of the invention. The cell may be transformed with the DNA construct of the invention encoding the variant, conveniently by integrating the DNA construct (in 20 one or more copies) in the host chromosome. This integration is generally considered to be an advantage as the DNA sequence is more likely to be stably maintained in the cell. Integration of the DNA constructs into the host chromosome may be performed according to conventional methods, e.g. by homologous or 25 heterologous recombination. Alternatively, the cell may be transformed with an expression vector as described above in connection with the different types of host cells.

The cell of the invention may be a cell of a higher organism such as a mammal or an insect, but is preferably a microbial cell, e.g. a bacterial or a fungal (including yeast) cell.

Examples of suitable bacteria are grampositive bacteria such as Bacillus subtilis, Bacillus licheniformis, Bacillus lentus, 35 Bacillus brevis, Bacillus stearothermophilus, Bacillus alkalophilus, Bacillus amyloliquefaciens, Bacillus coagulans, Bacillus circulans, Bacillus lautus, Bacillus megaterium, Bacillus thuringiensis, or Streptomyces lividans or Streptomyces

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murinus, or gramnegative bacteria such as E.coli. The transformation of the bacteria may, for instance, be effected by protoplast transformation or by using competent cells in a manner known per se.

5

The yeast organism may favourably be selected from a species of Saccharomyces or Schizosaccharomyces, e.g. Saccharomyces cerevisiae. The filamentous fungus may advantageously belong to a species of Aspergillus, e.g. Aspergillus oryzae or Aspergilio lus niger. Fungal cells may be transformed by a process involving protoplast formation and transformation of the protoplasts followed by regeneration of the cell wall in a manner known per se. A suitable procedure for transformation of Aspergillus host cells is described in EP 238 023.

15

In a yet further aspect, the present invention relates to a method of producing an α -amylase variant of the invention, which method comprises cultivating a host cell as described above under conditions conducive to the production of the variant and recovering the variant from the cells and/or culture medium.

The medium used to cultivate the cells may be any conventional medium suitable for growing the host cell in question and 25 obtaining expression of the α -amylase variant of the invention. Suitable media are available from commercial suppliers or may be prepared according to published recipes (e.g. as described in catalogues of the American Type Culture Collection).

30 The α-amylase variant secreted from the host cells may conveniently be recovered from the culture medium by well-known procedures, including separating the cells from the medium by centrifugation or filtration, and precipitating proteinaceous components of the medium by means of a salt such as ammonium sulphate, followed by the use of chromatographic procedures such as ion exchange chromatography, affinity chromatography, or the like.

Industrial Applications

The α -amylase variants of this invention possesses valuable properties allowing for various industrial applications. In particular the enzyme variants finds potential applications as a component in washing, dishwashing and hard surface cleaning detergent compositions, but it may also be useful in the production of sweeteners and ethanol from starch and for textile desizing. Conditions for conventional starch converting processes and liquefaction and/or saccharification processes are described in for instance US Patent No. 3,912,590 and EP patent publications Nos. 252,730 and 63,909.

Production of sweetners from starch: A "traditional" process for conversion of starch to fructose syrups normally consists of three consecutive enzymatic processes, viz. a liquefaction process followed by a saccharification process and an isomerization process. During the liquefaction process, starch is degraded to dextrins by an α-amylase (e.g. Termamyl") at pH values between 5.5 and 6.2 and at temperatures of 95-160°C for 20 a period of approx. 2h. In order to ensure an optimal enzyme stability under these conditions, lmM of calcium is added (40 ppm free calcium ions).

After the liquefaction process the dextrins are converted into 25 dextrose by addition of a glucoamylase (e.g. AMG*) and a debranching enzyme, such as an isoamylase or a pullulanase (e.g. Promozyme*). Before this step the pH is reduced to a value below 4.5, maintaining the high temperature (above 95°C), and the liquefying α-amylase activity is denatured. The tem10 perature is lowered to 60°C, and glucoamylase and debranching enzyme are added. The saccharification process proceeds for 24-72 hours.

After the saccharification process the pH is increased to a 3s value in the range of 6-8, preferably pH 7.5, and the calcium is removed by ion exchange. The dextrose syrup is then converted into high fructose syrup using, e.g., an immmobilized alucoseisomerase (such as Sweetzyme*).

35

At least 3 enzymatic improvements of this process could be obtained. All three improvements could be seen as individual benefits, but any combination (e.g. 1+2, 1+3, 2+3 or 1+2+3) could be employed:

<u>Improvement 1</u>. Reduction of the calcium dependency of the liquefying alpha-amylase.

Addition of free calcium is required to ensure adequately high stability of the α -amylase, but free calcium strongly inhibits the activity of the glucoseisomerase and needs to be removed, by means of an expensive unit operation, to an extent which reduces the level of free calcium to below 3-5 ppm. Cost savings could be obtained if such an operation could be avoided and the liquefaction process could be performed without addition of free calcium ions.

To achieve that, a less calcium-dependent Termamyl-like α -amylase which is stable and highly active at low 20 concentrations of free calcium (< 40 ppm) is required. Such a Termamyl-like α -amylase should have a pH optimum at a pH in the range of 4.5-6.5, preferably in the range of 4.5-5.5.

Improvement 2. Reduction of formation of unwanted Maillard
25 products

The extent of formation of unwanted Maillard products during the liquefaction process is dependent on the pH. Low pH favours reduced formation of Maillard products. It would thus be desirable to be able to lower the process pH from around pH 6.0 to a value around pH 4.5; unfortunately, all commonly known, thermostable Termamyl-like α -amylases are not very stable at low pH (i.e. pH < 6.0) and their specific activity is generally low.

Achievement of the above-mentioned goal requires a Termamyllike α -amylase which is stable at low pH in the range of

4.5-5.5 and at free calcium concentrations in the range of 0-40 ppm, and which maintains a high specific activity.

Improvement 3.

It has been reported previously (US patent 5,234,823) that when saccharifying with A. niger glucoamylase and B. acidopullulyticus pullulanase, the presence of residual α -amylase activity from the liquefaction process can lead to lower yields of dextrose if the α -amylase is not inactivated before the saccharification stage. This inactivation can typically be carried out by adjusting the pH to below 4.3 at 95°C, before lowering the temperature to 60°C for saccharification.

- 15 The reason for this negative effect on dextrose yield is not fully understood, but it is assumed that the liquefying α-amylase (for example Termamyl 120 L from B. licheniformis) generates "limit dextrins" (which are poor substrates for B. acidopullulyticus pullulanase) by hydrolysing 1,4-alpha-20 glucosidic linkages close to and on both sides of the branching points in amylopectin. Hydrolysis of these limit dextrins by glucoamylase leads to a build-up of the trisaccharide panose,
- 25 The development of a thermostable α-amylase which does not suffer from this disadvantage would be a significant process improvement, as no separate inactivation step would be required.

which is only slowly hydrolysed by glucoamylase.

30 If a Termamyl-like, low-pH-stable α -amylase is developed, an alteration of the specificity could be an advantage needed in combination with increased stability at low pH.

The methodology and principles of the present invention make it spossible to design and produce variants according to the invention having the required properties as outlined above.

Detergent Compositions

According to the invention, the α -amylase may typically be a component of a detergent composition. As such, it may be included in the detergent composition in the form of a nondusting granulate, a stabilized liquid, or a protected enzyme. 5 Non-dusting granulates may be produced, e.g. as disclosed in US 4,106,991 and 4,661,452 (both to Novo Industri A/S) and may optionally be coated by methods known in the art. Examples of waxy coating materials are poly(ethylene oxide) products (polyethyleneglycol, PEG) with mean molar weights of 1000 to 10 20000, ethoxylated nonylphenols having from 16 to 50 ethylene oxide units; ethoxylated fatty alcohols in which the alcohol contains from 12 to 20 carbon atoms and in which there are 15 to 80 ethylene oxide units; fatty alcohols; fatty acids; and mono- and di- and triglycerides of fatty acids. Examples of 15 film-forming coating materials suitable for application by fluid bed techniques are given in patent GB 1483591. Liquid enzyme preparations may, for instance, be stabilized by adding a polvol such as propylene glycol, a sugar or sugar alcohol, lactic acid or boric acid according to established methods. 20 Other enzyme stabilizers are well known in the art. Protected enzymes may be prepared according to the method disclosed in EP 238.216. The detergent composition of the invention may be in any convenient form, e.g. as powder, granules, paste or liquid. A 25 liquid detergent may be aqueous, typically containing up to 70%

The detergent composition comprises one or more surfactants, each of which may be anionic, nonionic, cationic, or zwitterionic. The detergent will usually contain 0-50% of anionic surfactant such as linear alkylbenzenesulfonate (LAS), alphaolefinsulfonate (LAS), alkyl sulfate (fatty alcohol sulfate) (As), alcohol ethoxysulfate (AEOS or AES), secondary alkanesulfonates (SAS), alpha-sulfo fatty acid methyl esters, alkylor alkenylsuccinic acid or soap. It may also contain 0-40% of nonionic surfactant such as alcohol ethoxylate (AEO or AE), carboxylated alcohol ethoxylates, nonylphenol ethoxylate, alkylpolyglycoside, alkyldimethylamineoxide, ethoxylated fatty

of water and 0-30% of organic solvent, or nonaqueous.

acid monoethanolamide, fatty acid monoethanolamide, or polyhydroxy alkyl fatty acid amide (e.g. as described in WO 92/06154).

5 The detergent composition may additionally comprise one or more other enzymes, such as lipase, cutinase, protease, cellulase, peroxidase, e.g., laccase.

The detergent may contain 1-65% of a detergent builder or complexing agent such as zeolite, diphosphate, triphosphate, phosphonate, citrate, nitrilotriacetic acid (NTA), ethylene-diaminetetraacetic acid (EDTA), diethylenetriaminepentaacetic acid (DTMPA), alkyl- or alkenylsuccinic acid, soluble silicates or layered silicates (e.g. SKS-6 from Hoechst). The 15 detergent may also be unbuilt, i.e. essentially free of detergent builder.

The detergent may comprise one or more polymers. Examples are carboxymethylcellulose (CMC), poly(vinylpyrrolidone) (PVP), 20 polyethyleneglycol (PEG), poly(vinyl alcohol) (PVA), polycarboxylates such as polyacrylates, maleic/acrylic acid copolymers and lauryl methacrylate/acrylic acid copolymers.

The detergent may contain a bleaching system which may comprise
25 a H₂O₂ source such as perborate or percarbonate which may be
combined with a peracid-forming bleach activator such as
tetraacetylethylenediamine (TAED) or nonanoyloxybenzenesulfonate (NOBS). Alternatively, the bleaching system may
comprise peroxy acids of e.g. the amide, imide, or sulfone
30 type.

The enzymes of the detergent composition of the invention may be stabilized using conventional stabilizing agents, e.g. a polyol such as propylene glycol or glycerol, a sugar or sugar 35 alcohol, lactic acid, boric acid, or a boric acid derivative as e.g. an aromatic borate ester, and the composition may be formulated as described in e.g. WO 92/19709 and WO 92/19708.

The detergent may also contain other conventional detergent ingredients such as e.g. fabric conditioners including clays, foam boosters, suds suppressors, anti-corrosion agents, soil-suspending agents, anti-soil redeposition agents, dyes, bactericides, optical brighteners, or perfume.

The pH (measured in aqueous solution at use concentration) will usually be neutral or alkaline, e.g. 7-11.

10 Particular forms of detergent compositions within the scope of the invention include:

1) A detergent composition formulated as a granulate having a 15 bulk density of at least 600 g/l comprising

	Linear alkylbenzenesulfonate (cal- culated as acid)	7	-	12%	
20	Alcohol ethoxysulfate (e.g. C ₁₂₋₁₈ alcohol, 1-2 EO) or alkyl sulfate (e.g. C ₁₆₋₁₈)	1	_	4%	
	Alcohol ethoxylate (e.g. C_{14-15} alcohol, 7 EO)	5	-	9%	
	Sodium carbonate (as Na ₂ CO ₃)	14		20%	
25	Soluble silicate (as Na20,2SiO2)	2	-	6%	
	Zeolite (as NaAlSiO4)	15	-	22%	
	Sodium sulfate (as Na ₂ SO ₄)	0	_	6%	
	Sodium citrate/citric acid (as $C_6H_5Na_3O_7/C_6H_8O_7$)	0		15%	
30	Sodium perborate (as NaBO ₃ .H ₂ O)	11		18%	
	TAED	2		6%	
	Carboxymethylcellulose	0	-	2%	
	Polymers (e.g. maleic/acrylic acid copolymer, PVP, PEG)	0	_	3 %	
35	Enzymes (calculated as pure enzyme protein)	٥.	0001	- 0.1%	
	Minor ingredients (e.g. suds suppressors, perfume, optical brightener, photobleach)	0	-	5%	

2) A detergent composition formulated as a granulate having a bulk density of at least $600~{\rm g/l}$ comprising

5	Linear alkylbenzenesulfonate (cal- culated as acid)	6 - 11%
	Alcohol ethoxysulfate (e.g. C ₁₂₋₁₈ alcohol, 1-2 EO or alkyl sulfate (e.g. C ₁₆₋₁₈)	1 - 3%
10	Alcohol ethoxylate (e.g. C_{14-15} alcohol, 7 EO)	5 - 9%
	Sodium carbonate (as Na ₂ CO ₃)	15 - 21%
	Soluble silicate (as Na ₂ O, 2SiO ₂)	1 - 4%
15	Zeolite (as NaAlSiO4)	24 - 34%
	Sodium sulfate (as Na ₂ SO ₄)	4 - 10%
	Sodium citrate/citric acid (as C _e H ₅ Na ₃ O ₃ /C _e H ₅ O ₃)	0 - 15%
	Carboxymethylcellulose	0 - 2%
20	Polymers (e.g. maleic/acrylic acid copolymer, PVP, PEG)	1 - 6%
	Enzymes (calculated as pure enzyme protein)	0.0001 - 0.1%
25	Minor ingredients (e.g. suds suppressors, perfume)	0 - 5%

3) A detergent composition formulated as a granulate having a bulk density of at least $600~{
m g/l}$ comprising

Linear alkylbenzenesulfonate (cal- culated as acid)	5	- 9%
Alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, 7 EO)	7	- 14%
Soap as fatty acid (e.g. C_{16-22} fatty acid)	1	- 3%
Sodium carbonate (as Na ₂ CO ₃)	10	- 17%
Soluble silicate (as Na ₂ O,2SiO ₂)	3	- 9%
Zeolite (as NaAlSiO4)	23	- 33%
Sodium sulfate (as Na ₂ SO4)	0	- 4%
Sodium perborate (as NaBO3.H2O)	8	- 16%
	culated as acid) Alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, 7 EO) Soap as fatty acid (e.g. C ₁₆₋₂₂ fatty acid) Sodium carbonate (as Na ₂ CO ₃) Soluble silicate (as Na ₂ O ₂ 2SiO ₂) Zeolite (as NaAlSiO ₄) Sodium sulfate (as Na ₂ SO4)	culated as acid) Alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, 7 7 EO) Soap as fatty acid (e.g. C ₁₆₋₂₂ fatty 1 acid) Sodium carbonate (as Na ₂ CO ₂) 10 Soluble silicate (as Na ₂ O ₂ SiO ₂) 3 Zeolite (as NaAlSiO ₄) 23 Sodium sulfate (as Na ₂ SO4) 0

TAED	2		8%
Phosphonate (e.g. EDTMPA)	0	_	1%
Carboxymethylcellulose	0	-	2%
Polymers (e.g. maleic/acrylic acid copolymer, PVP, PEG)	0	_	3%
Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
Minor ingredients (e.g. suds suppressors, perfume, optical brightener)	0	-	5%

4) A detergent composition formulated as a granulate having a bulk density of at least 600 g/l comprising

			_	
15	Linear alkylbenzenesulfonate (cal- culated as acid)	8	-	12%
	Alcohol ethoxylate (e.g. C_{12-15} alcohol, 7 EO)	10	-	25%
20	Sodium carbonate (as Na ₂ CO ₃)	14	-	22%
	Soluble silicate (as Na ₂ O, 2SiO ₂)	1	-	5%
	Zeolite (as NaAlSiO4)	25	-	35%
	Sodium sulfate (as Na2SO4)	0	-	10%
	Carboxymethylcellulose	0	-	2%
25	Polymers (e.g. maleic/acrylic acid copolymer, PVP, PEG)	1	_	3%
	Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
30	Minor ingredients (e.g. suds suppressors, perfume)	0	-	5%

5) An aqueous liquid detergent composition comprising

	Linear alkylbenzenesulfonate (cal- culated as acid)	15	- 21%	
35	Alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, hol, 7 EO or C ₁₂₋₁₅ alcohol, 5 EO)	12	- 18%	
	Soap as fatty acid (e.g. oleic acid)	3	- 13%	
40	Alkenylsuccinic acid (C12-14)	0	- 13%	

Aminoethanol	8 - 18%
Citric acid	2 - 8%
Phosphonate	0 - 3%
Polymers (e.g. PVP, PEG)	0 - 3%
Borate (as B ₄ O ₇)	0 - 2%
Ethanol	0 - 3%
Propylene glycol	8 - 14%
Enzymes (calculated as pure enzyme protein)	0.0001 - 0.1%
Minor ingredients (e.g. dispersants, suds suppressors, perfume, optical brightener)	0 - 5%

6) An aqueous structured liquid detergent composition comprising in ${\bf g}$

			-	
	Linear alkylbenzenesulfonate (calculated as acid)	15		21%
20	Alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, 7 EO, or C ₁₂₋₁₅ alcohol, 5 EO)	3	-	9%
	Soap as fatty acid (e.g. oleic acid)	3	-	10%
	Zeolite (as NaA1SiO ₄)	14	-	22%
	Potassium citrate	9	_	18%
25	Borate (as B ₄ O ₇)	0	-	2%
	Carboxymethylcellulose	0	-	2*
	Polymers (e.g. PEG, PVP)	0	-	3%
30	Anchoring polymers such as, e.g., lauryl methacrylate/acrylic acid copolymer; molar ratio 25:1; MW 3800	0	-	3 %
	Glycerol	0	-	5%
	Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
35	Minor ingredients (e.g. dispersants, suds suppressors, perfume, optical brighteners)	o	-	5%

7) A detergent composition formulated as a granulate having a bulk density of at least 600 g/l comprising

	Fatty alcohol sulfate	5	_	10%
5	Ethoxylated fatty acid monoethanol-amide	3	-	9%
	Soap as fatty acid	0	-	3 %
	Sodium carbonate (as Na ₂ CO ₃)	5	-	10%
	Soluble silicate (as Na ₂ O, 2SiO ₂)	1	-	4%
	Zeolite (as NaAlSiO4)	20	-	40%
10	Sodium sulfate (as Na ₂ SO ₄)	2	-	8%
	Sodium perborate (as NaBO3.H2O)	12	-	18%
	TAED	2	-	7%
	Polymers (e.g. maleic/acrylic acid copolymer, PEG)	1	-	5%
15	Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
	Minor ingredients (e.g. optical brightener, suds suppressors, per- fume)	0	-	5%

8) A detergent composition formulated as a granulate comprising

	Linear alkylbenzenesulfonate (calculated as acid)	8	_	14%
25	Ethoxylated fatty acid monoethanol-amide	5	-	11%
	Soap as fatty acid	0	-	3%
	Sodium carbonate (as Na ₂ CO ₃)	4	-	10%
	Soluble silicate (as Na ₂ O, 2SiO ₂)	1	-	4%
	Zeolite (as NaAlSiO4)	30	-	50%
0	Sodium sulfate (as Na ₂ SO ₄)	3	-	11%
	Sodium citrate (as C ₆ H ₅ Na ₃ O ₇)	5	-	12%
	Polymers (e.g. PVP, maleic/acrylic acid copolymer, PEG)	1	-	5%
5	Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
	Minor ingredients (e.g. suds suppressors, perfume)	0	-	5%

9) A detergent composition formulated as a granulate comprising

	Linear alkylbenzenesulfonate (calculated as acid)	6	_	12%
	Nonionic surfactant	1	-	4%
5	Soap as fatty acid	2	-	6%
	Sodium carbonate (as Na ₂ CO ₃)	14	-	22%
	Zeolite (as NaAlSiO,)	18	-	32%
	Sodium sulfate (as Na ₂ SO ₄)	5	-	20%
	Sodium citrate (as C ₆ H ₅ Na ₃ O ₇)	3	-	8%
10	Sodium perborate (as NaBO3.H2O)	4	-	9%
	Bleach activator (e.g. NOBS or TAED)	. 1	-	5%
	Carboxymethylcellulose	0	_	2%
15	Polymers (e.g. polycarboxylate or PEG)	1	-	5%
	Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
	Minor ingredients (e.g. optical brightener, perfume)	0	-	5%
			_	

10) An aqueous liquid detergent composition comprising

Linear alkylbenzenesulfonate (calculated as acid)	15	- 23%	
Alcohol ethoxysulfate (e.g. C ₁₂₋₁₅ alcohol, 2-3 EO)	8	- 15%	
Alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, 7 EO, or C ₁₂₋₁₅ alcohol, 5 EO)	3	- 9%	
Soap as fatty acid (e.g. lauric acid)	0	- 3%	
Aminoethanol	1	- 5%	_
Sodium citrate	5	- 10%	
Hydrotrope (e.g. sodium toluensulfonate)	2	- 6%	
Borate (as B ₄ O ₇)	0	- 2%	
Carboxymethylcellulose	0	- 1%	
Ethanol	1	- 3%	
Propylene glycol	2	- 5%	
	(calculated as acid) Alcohol ethoxysulfate (e.g. C ₁₂₋₁₅ alcohol, 2-3 EO) Alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, 7 EO, or C ₁₂₋₁₅ alcohol, 5 EO) Soap as fatty acid (e.g. lauric acid) Aminoethanol Sodium citrate Hydrotrope (e.g. sodium toluensulfonate) Borate (as B ₄ O ₇) Carboxymethylcellulose	(calculated as acid) 15 Alcohol ethoxysulfate (e.g. C ₁₂₋₁₅ alcohol, 2-3 EO) 8 Alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, 7 EO, or C ₁₂₋₁₅ alcohol, 5 EO) 3 Soap as fatty acid (e.g. lauric acid) 0 Aminoethanol 1 Sodium citrate 5 Hydrotrope (e.g. sodium toluensulfonate) 2 Borate (as B ₄ O ₂) 0 Carboxymethylcellulose 0 Ethanol 1	(calculated as acid) 15 - 23% Alcohol ethoxysulfate (e.g. C ₁₂₋₁₅ alcohol, 2-3 EO) 8 - 15% Alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, 7 EO, or C ₁₂₋₁₅ alcohol, 5 3 - 9% EO) Soap as fatty acid (e.g. lauric acid) 0 - 3% Aminoethanol 1 - 5% Sodium citrate 5 - 10% Hydrotrope (e.g. sodium toluensulfonate) 2 - 6% Borate (as E ₄ O ₇) 0 - 2% Carboxymethylcellulose 0 - 1% Ethanol 1 - 3%

Enzymes (calculated as pure enzyme protein)	0.0001 -	0.1%	
Minor ingredients (e.g. polymers, dispersants, perfume, optical brighteners)	0 -	5%	

11) An aqueous liquid detergent composition comprising

	Linear alkylbenzenesulfonate (calculated as acid)	20	_	32%
10	Alcohol ethoxylate (e.g. C_{12-15} alcohol, 7 EO, or C_{12-15} alcohol, 5 EO)	6	-	12%
	Aminoethanol	2	-	6%
	Citric acid	8	-	14%
15	Borate (as B ₄ O ₇)	1	-	3%
20	Polymer (e.g. maleic/acrylic acid copolymer, anchoring polymer such as, e.g., lauryl methacrylate/acrylic acid copolymer)	0	-	3%
	Glycerol	3	-	8%
	Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
25	Minor ingredients (e.g. hydro- tropes, dispersants, perfume, optical brighteners)	0	-	5%

12) A detergent composition formulated as a granulate having a bulk density of at least 600 g/l comprising

30	Anionic surfactant (linear alkylbenzenesulfonate, alkyl sulfate, alpha-olefinsulfonate, alphasulfo fatty acid methyl esters, alkanesulfonates, soap)	25	- 40%
35	Nonionic surfactant (e.g. alcohol ethoxylate)	1	- 10%
	Sodium carbonate (as Na ₂ CO ₃)	8	- 25%
	Soluble silicates (as Na ₂ O, 2SiO ₂)	5	- 15%
	Sodium sulfate (as Na ₂ SO ₄)	0	- 5%
40	Zeolite (as NaA1SiO4)	15	- 28%
	Sodium perborate (as NaBO, .4H2O)	0	- 20%

Bleach activator (TAED or NOBS)	0	-	5%
Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
Minor ingredients (e.g. perfume, optical brighteners)	0	-	3%

13) Detergent formulations as described in 1) - 12) wherein all or part of the linear alkylbenzenesulfonate is replaced by $(C_{12}-C_{14})$ alkyl sulfate.

10

14) A detergent composition formulated as a granulate having a bulk density of at least 600 g/l comprising

	(C ₁₂ -C ₁₈) alkyl sulfate	9		15%
15	Alcohol ethoxylate	3	-	6%
	Polyhydroxy alkyl fatty acid amide	1	-	5%
	Zeolite (as NaA1SiO4)	10	-	20%
	Layered disilicate (e.g. SK56 from Hoechst)	10	_	20%
20	Sodium carbonate (as Na ₂ CO ₃)	3	_	12%
	Soluble silicate (as Na ₂ O, 2SiO ₂)	0	-	6%
	Sodium citrate	4	-	8%
	Sodium percarbonate	13	-	22%
	TAED	3	_	8%
25	Polymers (e.g. polycarboxylates and PVP=	0	-	5%
	Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
30	Minor ingredients (e.g. optical brightener, photo bleach, perfume, suds suppressors)	0	-	5%

15) A detergent composition formulated as a granulate having $_{35}$ a bulk density of at least 600 g/l comprising

(C,2-C,e) alkyl sulfate	4	- 8%
Alcohol ethoxylate	11	- 15%

Soap	1		4%
Zeolite MAP or zeolite A	35	-	45%
Sodium carbonate (as Na ₂ CO ₃)	2	-	8%
Soluble silicate (as Na ₂ O,2SiO ₂)	0	-	4%
Sodium percarbonate	13	-	22%
TAED	1	-	8%
Carboxymethyl cellulose	0	_	3%
Polymers (e.g. polycarboxylates and PVP)	0	-	3%
Enzymes (calculated as pure enzyme protein)	0.0001	-	0.1%
Minor ingredients (e.g. optical brightener, phosphonate, perfume)	. 0	-	3%

- 15 16) Detergent formulations as described in 1) 15) which contain a stabilized or encapsulated peracid, either as an additional component or as a substitute for already specified bleach systems.
- 20 17) Detergent compositions as described in 1), 3), 7), 9) and
 - 12) wherein perborate is replaced by percarbonate.
 - 18) Detergent compositions as described in 1), 3), 7), 9), 12),
 - 14) and 15) which additionally contain a manganese catalyst.
- 25 The manganese catalyst may, e.g., be one of the compounds described in "Efficient manganese catalysts for low-temperature bleaching", Nature 362, 1994, pp. 637-639.
 - 19) Detergent composition formulated as a nonaqueous detergent
- 30 liquid comprising a liquid nonionic surfactant such as, e.g., linear alkoxylated primary alcohol, a builder system (e.g. phosphate), enzyme and alkali. The detergent may also comprise anionic surfactant and/or a bleach system.
- 35 The α-amylase variant of the invention may be incorporated in concentrations conventionally employed in detergents. It is at present contemplated that, in the detergent composition of the invention, the α-amylase may be added in an amount correspon-

ding to 0.00001-1 mg (calculated as pure enzyme protein) of α -amylase per liter of wash liquor.

Dishwashing Composition

- 5 The dishwashing detergent composition comprises a surfactant which may be anionic, non-ionic, cationic, amphoteric or a mixture of these types. The detergent will contain 0-90% of non-ionic surfactant such as low- to non-foaming ethoxylated propoxylated straight-chain alcohols.
- The detergent composition may contain detergent builder salts of inorganic and/or organic types. The detergent builders may be subdivided into phosphorus-containing and non-phosphorus-containing types. The detergent composition usually contains 1-90% of detergent builders.
- Examples of phosphorus-containing inorganic alkaline detergent builders, when present, include the water-soluble salts especially alkali metal pyrophosphates, orthophosphates, and polyphosphates. An example of phosphorus-containing organic alkaline detergent builder, when present, includes the water-
- 20 soluble salts of phosphonates. Examples of non-phosphoruscontaining inorganic builders, when present, include watersoluble alkali metal carbonates, borates and silicates as well as the various types of water-insoluble crystalline or amorphous alumino silicates of which zeolites are the best-known
- 25 representatives.

Examples of suitable organic builders include the alkali metal, ammonium and substituted ammonium, citrates, succinates, malonates, fatty acid sulphonates, carboxymetoxy succinates, an ammonium polyacetates, carboxylates, polycarboxylates, aminopolycarboxylates, polyacetyl carboxylates and polyhydroxsulphonates.

Other suitable organic builders include the higher molecular 35 weight polymers and co-polymers known to have builder properties, for example appropriate polyacrylic acid, polymaleic and polyacrylic/polymaleic acid copolymers and their salts.

The dishwashing detergent composition may contain bleaching agents of the chlorine/bromine-type or the oxygen-type. Examples of inorganic chlorine/bromine-type bleaches are lithium, sodium or calcium hypochlorite and hypobromite as wells as chlorinated trisodium phosphate. Examples of organic chlorine/bromine-type bleaches are heterocyclic N-bromo and N-chloro imides such as trichloroisocyanuric, tribromoisocyanuric, dibromoisocyanuric and dichloroisocyanuric acids, and salts thereof with water-solubilizing cations such as potassium and sodium. Hydantoin compounds are also suitable.

The oxygen bleaches are preferred, for example in the form of an inorganic persalt, preferably with a bleach precursor or as a peroxy acid compound. Typical examples of suitable peroxy is bleach compounds are alkali metal perborates, both tetrahydrates and monohydrates, alkali metal percarbonates, persilicates and perphosphates. Preferred activator materials are TAED and diverol triacetate.

20 The dishwashing detergent composition of the invention may be stabilized using conventional stabilizing agents for the enzyme(s), e.g. a polyol such as e.g.propylene glycol, a sugar or a sugar alcohol, lactic acid, boric acid, or a boric acid derivative, e.g. an aromatic borate ester.

25

The dishwashing detergent composition of the invention may also contain other conventional detergent ingredients, e.g. deflocculant material, filler material, foam depressors, anti-corrosion agents, soil-suspending agents, sequestering agents, anti-soil redeposition agents, dehydrating agents, dyes, bactericides, fluorescers, thickeners and perfumes.

Finally, the α -amylase variant of the invention may be used in conventional dishwashing detergents, e.g. in any of the detergents described in any of the following patent publications:

EP 518719, EP 518720, EP 518721, EP 516553, EP 516554,

EP 516555, GB 2200132, DE 3741617, DE 3727911, DE 4212166, DE 4137470, DE 3833047, WO 93/17089, DE 4205071, WO 52/09680, WO 93/18129, WO 93/04153, WO 92/06157, WO 92/08777, EP 429124, WO 93/21299, US 5141664, EP 561452, EP 561446, GB 2234980, 5 WO 93/03129, EP 481547, EP 530870, EP 533239, EP 554943, EP 346137, US 5112518, EP 318204, EP 318279, EP 271155, EP 271156, EP 346136, GB 2228945, CA 2006687, WO 93/25651, EP 530635, EP 414197, US 5240632.

10 EXAMPLES

EXAMPLE 1

15 Example on Homology building of TERM

The overall homology of the B. licheniformis α-amylase (in the following referred to as TERM) to other Termamyl-like α-amylases is high and the percent similarity is extremely high.

The similarity calculated for TERM to BSG (the B. stearothermophilus α-amylase with SEQ ID NO 6), and BAN (the B. amyloliquefaciens α-amylase with SEQ ID NO 4) using the University of Wisconsin Genetics Computer Group's program GCG gave 89% and 78%, respectively. TERM has a deletion of 2 residues between residue G180 and K181 compared to BAN and BSG. BSG has a deletion of 3 residues between G371 and I372 in comparison with BAN and TERM. Further BSG has a C-terminal extension of more than 20 residues compared to BAN and TERM. BAN has 2 residues less and TERM has one residue less in the N-terminal compared to BSG.

The structure of the B. licheniformis (TERM) and of the B. amyloliquefaciens α -amylase (BAN), respectively, was model built on the structure disclosed in Appendix 1 herein. The structure of other Termamyl-like α -amylases (e.g. those disclosed herein) may be built analogously.

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In comparison with the α-amylase used for elucidating the present structure, TERM differs in that it lacks two residues around 178-182. In order to compensate for this in the model structure, the HOMOLOGY program from BIOSYM was used to substitute the residues in equivalent positions in the structure (not only structurally conserved regions) except for the deletion point. A peptide bond was established between G179(G177) and K180(K180) in TERM(BAN). The close structural relationship between the solved structure and the model structure (and thus the validity of the latter) is indicated by the presence of only very few atoms found to be too close together in the model.

To this very rough structure of TERM was then added all waters 15 (605) and ions (4 Calcium and 1 Sodium) from the solved structure (Appendix 1) at the same coordinates as for said solved structure using the INSIGHT program. This could be done with only few overlaps - in other words with a very nice fit. This model structure were then minimized using 200 steps of 20 Steepest descent and 600 steps of Conjugated gradient (see Brooks et al 1983, J. Computational Chemistry 4, p.187-217). The minimized structure was then subjected to molecular dynamics, 5ps heating followed by up to 200ps equilibration but more than 35ps. The dynamics as run with the Verlet algorithm 25 and the equilibration temperature 300K were kept using the Behrendsen coupling to a waterbath (Berendsen et. al., 1984, J. Chemical Physics 81, p. 3684-3690). Rotations and translations were removed every picosecond. The potential energy became stable after appr. 35ps equilibration. A mean dynamics struc-30 ture was extracted and can be used for further analysis.

EXAMPLE 2

Determination of residues within 10Å from the ions present in 35 the solved structure

The coordinates of Appendix 1 are read into the INSIGHT program provided by BIOSYM tecnologies. The spatial coordinates are

presented showing the bonds between the atoms. The ions are presented as well as the water atoms. The program package part of creating subset are used to create a 10Å subset around the Calcium and the Sodium ions in the structure using the command 5 ZONE. All residues having an atom within the 10Å are compiled and written out by the LIST MOLECULE command. By giving the ions the name ium in the coordinate file a 10Å sphere around all atoms called ium is compiled. The specific residues identified in this manner are given further above in the 10 section entitled "Ca² dependency".

EXAMPLE 3

WO 96/23874

Determination of cavities in the solved structure (Appendix 1)

The solved structure exhibits many internal holes and cavities. When analysing for such cavities the Connolly program is normally used (Lee, B. and Richards, F.M. (1971) J. Mol. Biol. 55,p. 379-400). The program uses a probe with radius to search the external and internal surface of the protein. The smallest hole observable in this way has the probe radius.

To analyse the solved structure a modified version of the Connolly program included in the program of INSIGHT were used.

25 First the water molecules and the ions were removed by unmerging these atoms from the solved structure. By using the command MOLECULE SURFACE SOLVENT the solvent accessible surface area were calculated for all atoms and residues using a probe radius of 1.4Å, and displayed on the graphics screen together with the model of the solved structure. The internal cavities where then seen as dot surfaces with no connections to external surface.

Mutant suggestions for filling out the holes are given in the specification (in the section entitled "Variants with increased thermostability and/or altered temperature optimum"). By using the homology build structures or/and the sequence alignment

mutations for the homologous structures of TERM and BSG and ${\tt BAN}$ can be made.

EXAMPLE 4

Construction of Termamyl m variants in accordance with the invention

Termamyl (SEQ ID NO. 2) is expressed in B. subtilis from a plasmid denoted pDN1528. This plasmid contains the complete gene encoding Termamyl, amyL, the expression of which is directed by its own promoter. Further, the plasmid contains the origin of replication, ori, from plasmid pUB110 and the cat gene from plasmid pC194 conferring resistance towards to chloramphenicol. pDN1528 is shown in Fig. 9.

A specific mutagenesis vector containing a major part of the coding region of SEQ ID NO 1 was prepared. The important features of this vector, denoted pJeEN1, include an origin of replication derived from the pUC plasmids, the cat gene conferring resistance towards chloramphenicol, and a frameshift-containing version of the bla gene, the wild type of which normally confers resistance towards ampicillin (amp⁸ phenotype). This mutated version results in an amp⁸ phenotype.

The plasmid pJeEN1 is shown in Fig. 10, and the E. coli origin of replication, ori, bla, cat, the 5'-truncated version of the Termamyl amylase gene, and selected restriction sites are indicated on the plasmid.

Nutations are introduced in amyL by the method described by Deng and Nickoloff (1992, Anal. Biochem. 200, pp. 81-88) except that plasmids with the "selection primer" (primer #6616; see below) incorporated are selected based on the amp^R phenotype of transformed E. coli cells harboring a plasmid with a repaired bla gene, instead of employing the selection by restriction enzyme digestion outlined by Deng and Nickoloff. Chemicals and enzymes used for the mutagenesis were obtained from the

Chameleon $^{\mathbf{m}}$ mutagenesis kit from Stratagene (catalogue number 200509).

After verification of the DNA sequence in variant plasmids, the struncated gene, containing the desired alteration, is subcloned into pDN1528 as a PstI-EcoRI fragment and transformed into a protease- and amylase-depleted Bacillus subtilis strain in order to express the variant enzyme.

10 The Termamyl variant V54W was constructed by the use of the following mutagenesis primer (written 5' to 3', left to right):

PG GTC GTA GGC ACC GTA GCC CCA ATC CGC TTG

15 The Termamyl variant A52W + V54W was constructed by the use of the following mutagenesis primer (written 5' to 3', left to right):

PG GTC GTA GGC ACC GTA GCC CCA ATC CCA TTG GCT CG

20

Primer #6616 (written 5' to 3', left to right; P denotes a 5' phosphate):

P CTG TGA CTG GTG AGT ACT CAA CCA AGT C

25

EXAMPLE 5

Saccharification in the presence of "residual" α -amylase activity

30

Two appropriate Termamyl variants with altered specificity were evaluated by saccharifying a DE 10 (DE = dextrose equivalent) maltodextrin substrate with A. niger glucoamylase and B. acidopullulyticus pullulanase under conditions where the variant amylase was active.

Saccharification: Substrates for saccharification were prepared by dissolving 230 g DE 10 spray-dried maltodextrin, prepared

10

from common corn starch, in 460 ml boiling deionized water and adjusting the dry substance (DS) content to approximately 30% w/w. The pH was adjusted to 4.7 (measured at 60°C) and aliquots of substrate corresponding to 15 g dry weight were transferred to 50 ml blue cap glass flasks.

The flasks were then placed in a shaking water bath equilibrated at $60\,^{\circ}\text{C}$, and the enzymes added. The pH was readjusted to 4.7 where necessary.

The following enzymes were used:

Glucoamylase: AMG™ (Novo Nordisk A/S); dosage 0.18 AG/g DS

Pullulanase: Promozyme™ (Novo Nordisk A/S);

dosage 0.06 PUN/g DS

 α -Amylases: Termamyl m (Novo Nordisk A/S); dosage 60 NU/g DS Termamyl variant V54W; dosage 60 NU/g DS

Termamyl variant V54W + A52W; dosage 60 NU/g DS

20 2 ml samples were taken periodically. The pH of each sample was adjusted to about 3.0, and the sample was then heated in a boiling water bath for 15 minutes to inactivate the enzymes. After cooling, the samples were treated with approximately 0.1 g mixed-bed ion exchange resin (BIO-Rad 501-X (D)) for 30 25 minutes on a rotary mixer and then filtered. The carbohydrate composition of each sample was determined by HPLC. The following results were obtained after 72 hours [DP_n denotes a dextrose (D-glucose) oligomer with n glucose units]:

α -amylase	%DP ₁	%DP ₂	%DP ₃	%DP4
None (control)	95.9	2.8	0.4	1.0
V54W	96.0	2.9	0.4	0.8
V54W + A52W	95.9	2.8	0.4	0.8
Termamyl™	95.6	2.8	0.8	0.8

It can be seen from the above results that compared with the control (no α -amylase activity present during liquefaction), the presence of α -amylase activity from variants V54W and V54W + A52W did not lead to elevated panose (DP₃) levels. In 5 contrast, Termamyl α -amylase activity resulted in higher levels of panose and a subsequent loss of D-glucose (DP₁) yield.

Thus, if α -amylase variants V54W or V54W + λ 52W are used for starch liquefaction, it will not be necessary to inactivate the 10 residual α -amylase activity before the commencement of saccharification.

EXAMPLE 6

15 Calcium-binding affinity of α -amylase variants of the invention

Unfolding of amylases by exposure to heat or to denaturants such as guanidine hydrochloride is accompanied by a decrease in fluorescence. Loss of calcium ions leads to unfolding, and the affinity of α -amylases for calcium can be measured by fluorescence measurements before and after incubation of each α -amylase (e.g. at a concentration of 10 μ g/ml) in a buffer (e.g. 50 mM HEPES, pH 7) with different concentrations of calcium (e.g. in the range of 1 μ M-100 mM) or of EGTA (e.g. in 25 the range of 1-1000 μ M) [EGTA = 1,2-di(2-aminoethoxy)ethane-N, N', N' -tetraacetic acid] for a sufficiently long period of time (such as 22 hours at 55°C).

The measured fluorescence F is composed of contributions form
the folded and unfolded forms of the enzyme. The following
equation can be derived to describe the dependence of F on
calcium concentration ([Ca]):

$$F = [Ca] / (K_{diss} + [Ca]) (\alpha_{N} - \beta_{N} log([Ca])) + K_{diss} / (K_{diss} + [Ca]) (\alpha_{U} - \beta_{U} log([Ca]))$$

where α_N is the fluorescence of the native (folded) form of the enzyme, β_N is the linear dependence of α_N on the logarithm of

the calcium concentration (as observed experimentally), α_{0} is the fluorescence of the unfolded form and β_{0} is the linear dependence of α_{0} on the logarithm of the calcium concentration. K_{diss} is the apparent calcium-binding constant for an equilibrium 5 process as follows:

 K_{diss} N-Ca \leftrightarrow U + Ca (N = native enzyme; U = unfolded enzyme)

- 10 In fact, unfolding proceeds extremely slowly and is irreversible. The rate of unfolding is a dependent on calcium concentration, and the dependency for a given α-amylase provides a measure of the Ca-binding affinity of the enzyme. By defining a standard set of reaction conditions (e.g. 22 hours at 55°C), a meaningful comparison of K_{diss} for different α-amylases can be made. The calcium dissociation curves for α-amylases in general can be fitted to the equation above, allowing determination of the corresponding values of K_{tiss}.
- The following values for $K_{\rm diss}$ were obtained for a parent Termamyl-like α -amylase having the amino acid sequence shown in SEQ ID No. 1 of WO 95/26397 and for the indicated variant thereof according to the invention:

 $\frac{\text{25 } \alpha\text{-Amylase}}{\text{L351C} + \text{M430C} + \text{T183*} + \text{G184*} 1.7 \ (\pm 0.5) \ \times \ 10^{-3}}{\text{3.5 } (\pm 1.1) \ \times \ 10^{-1}}$

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It is apparent from the above that the calcium-binding affinity of the variant in question binds calcium significantly more strongly than the parent, and thereby has a correspondingly lower calcium dependency than the parent.

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SEQUENCE LISTING

In the following SEQ ID Nos. 1, 3, 5 the 5', coding sequence and 3' sequence of the relevant α-amylase genes are illustrated. The 5' sequence is the first separate part of the sequence written with lower case letters, the coding sequence is the intermediate part of the sequence, where the signal sequence is written with lower case letters and the sequence encoding the mature α-amylase is written with upper case letters, and to the 3' sequence is the third separate part of the sequence written with lower case letters.

SEQ ID No. 1

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cggaagattggaagtacaaaaataagcaaaagattgtcaatcatgtcatgagccatgcgggagacggaaaaatcgtctta atgcacgatatttatgcaacgttcgcagatgctgctgaagagattattaaaaagctgaaagcaaaaggctatcaattggt aactgtatctcagcttgaagaagtgaagaagcagagaggctattgaataaatgagtagaagcgccatatcggcgctttc

20 ttttggaagaaaatatagggaaaatggtacttgttaaaaattcggaatatttatacaacatcatatgtttcacattgaaa ggggaggagaacc

atgaaacaacaaaaacqqctttacqcccqattgctqacqctgttatttgcgctcatcttcttgctgc ctcattctqcaqcaqcqqcqGCAAATCTTAATGGGACGCTGATGCAGTATTTT-25 GAATGGTACATGCCCAATGACGGCCAA CATTGGAGGCGTTTGCAAAACGACTCGGCATAT-TTGGCTGAACACGGTATTACTGCCGTCTGGATTCCCCCGGCATATAA CAAGCGGATGTGGGCTACGGTGCTTACGACCTTTATGATTTAGGGGAGTTTCATCAAAAAG-GGACGGTTC GGACAAAGTACGGCACAAAAGGAGCTGCAATCTGCGATCAAAAGTCTTC-ATTCCCGCGACATTAACGTTTACGGGGAT GTGGTCATCAACCACAAAGGCGGCGCTGA-30 TGCGACCGAAGATGTAACCGCGGTTGAAGTCGATCCCGCTGACCGCCAACCG CGTAATTT-CAGCGATTTTA AATGGCATTGGTACCATTTTGACGGAACCGATTGGGACGAGTCCCGAAA-GCTGAACCGCATCTATAAGTTTCAAGGAAAG GCTTGGGATTGGGAAGTTTCCAATGAA-AACGGCAACTATGATTATTTGATGTATGCCGACATCGATTATGACCATCCTGA TGTCGCAG-35 CAGAAATTAAGAGATGGGGCACTTGGTATGCCAATGAACTGCAATTGGACGGTTTCCGTCTT-GATGCTGTCA AACACATTAAATTTTCTTTTTTGCGGGATTGGGTTAATCATGTCAGGGA-AAAAACGGGGAAGGAAATGTTTACGGTAGCT GAATATTGGCAGAAT -CACTTGGGCGCGCTGGAAAACTATTTGAACAAAACAAATTTTAATCATTCAGTGTTTGAC-

84

GTGCC GCTTCATTATCAGTTCCATGCTGCATCGACACAGGGAGGCGGCTATGATATGAG-GAAATTGCTGAACGGTACGGTCGTTT CCAAGCATCCGTTGAAATCGGTTACATTTGTCG-ATAACCATGATACACAGCCGGGGCAATCGCTTGAGTCGACTGTCCAA ACATGGTTTAAG-CCGCTTGCTTACGCTTTTATTCTCACAAGGGAATCTGGATACCCTCAGGTTTTCTACGGG-

10 ATTCGGAAGGCTGGGGAGAGTTTCACGTAAACG GCGGGTCGGTTTCAATTTATGTTCAAA-GATAG

aagagcagagaggacggatttcctgaaggaaatccgtttttttatttt

15 SEQ ID No. 2

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GELQSAIKSLHSRDINVYGDVVINHKGGADATEDVTAVEV
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DGTDWDESRKLNRIYKFQGKAWDWEVSNENGNYDYLMYAD
IDYDHPDVAAEIKRWGTWYANELQLDGFRLDAVKHIKFSF
LRDWVNHVREKTGKEMFTVAEYWQNDLGALENYLNKTNFN
HSVFDVPLHYQFHAASTQGGGYDMRKLLNGTVVSKHPLKS
VTFVDNHDTQPGQSLESTVQTWFKPLAYAFILTRESGYPQ
VFYGDMYGTKGDSQREIPALKHKIEPILKRKQYAYGAQH
DYFDHHDIVGWTREGDSSVANSGLAALITDGPGGAKRMYV
GRQNAGETWHDITGNRSEPVVINSEGWGEFHVNGGSVSIY
VOR

30

SEO ID No. 3

gccccgcacatacgaaaagactggctgaaaaacattgagcctttgatgactgatgattttgg-35 ctgaagaagtggatcgattg tttgagaaaaagaagaagaacataaaaataccttgtctgtcatcagacagggtatttttttatgctgtccagactgtccgct gtgtaaaaataaaggaataaagggggttgttattatttttactgatatgtaaaatataatttgtataagaaaatgagagg agaggaaaac

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atgattcaaaaagaaaggacagtttcgttcagacttgtgcttatgtgcacgctgttattgtcagttt gccgattacaaaaacatcagcGTAAATGGCACGCTGATGCAGTATTTTGAATGGTATACGCCGAACGACGCCACCAT GGAAACGATTGCAGAATGATGCGGAACATTTATCGGATACCGGAATCACTGCCGTCTGGATTCCTCCCCCATACAAAGGA TTGAG5 CCAATCCGATAACGGACCTTATGATTTGTATGATTTAGGAGAATTCCAGCAAAAAGGGACGTCAGAAC GAAATACGGACAAAATCAGAGCTTCAAGATGCGTCGCTCACTGCATTCCCGGAACGTCCAAGTATACGGAGATTGG TTTTGAATCATAAGGCTGGTGCT-

- 10 AGGTACAGTGATTTTAAATG GCATTGGTATCATTTCGAGGGAGCGGACTGGGATGAATCCC-GGAAGATCAGCCGCATCTTTAAGTTTCGTGGGGAAGGAA AAGCGTGGGATTGGGAAGTAT-CAAGTGAAAACGGCAACTATGACTATTTAATGTATGCTGATTGTGATCTACCACCCCC GATGTCGTGGCAGGAGCAAAAAAATGGGGTATCTGGTATGCGAATGAACTGCTCATTAGACGG-CTTCCGTATTGATGCCGC CAAACATATTAAATTTTCATTTCTGCGTGATTGGGTTCAGG-
- 20 ACCGCTTGCATACGCCTTTATTTTGACAAGAGAATCCGGTTATCCTCAGGTGTTCTATGGGGATATG TACGGGACAAAAGGGACATCGCCAAAGGAAATTCCCTCACTGAAAGATAATATAGAGCCGATTTTAAAAGCGCGTAAGGA GTACGCATACGGGCCCCAGCACGATTATATTGACCACCCGGATTGATCGGATGGACGAGGGAAGGTGACAGGTCCGCCG CCAAATCAGGTTTGGCCGCTTTAATCACGGACGGACCCGGCGGATCAAAGCGGATGTATGCCGG-
- 25 CCTGAAAAATGCCGGC GAGACATGGTATGACATAACGGGCAACCGTTCAGATACTGTAA-AAATCGGATCTGACGGCTGGGGAGAGTTTCATGTAAA CGATGGGTCCGTCTCCATTTAT-GTTCAGAAATAA

86

SEQ ID No. 4

VNGTLMQYFEWYTPNDGQHWKRLQNDAEHLSDIGITAVWI PPAYKGLSOSDNGYGPYDLYDLGEFQQKGTVRTKYGTKSE 5 LODAIGSLHSRNVOVYGDVVLNHKAGADATEDVTAVEVNP ANRNOETSEEYOIKAWTDFRFPGRGNTYSDFKWHWYHFDG ADWDESRKISRIFKFRGEGKAWDWEVSSENGNYDYLMYAD VDYDHPDVVAETKKWGIWYANELSLDGFRIDAAKHIKESF LRDWVQAVRQATGKEMFTVAEYWQNNAGKLENYLNKTSFN 10 OSVFDVPLHFNLOAASSOGGGYDMRRLLDGTVVSRHPEKA VTFVENHDTQPGOSLESTVOTWFKPLAYAFILTRESGYPO VFYGDMYGTKGTSPKEIPSLKDNIEPILKARKEYAYGPOH DYIDHPDVIGWTREGDSSAAKSGLAALITDGPGGSKRMYA GLKNAGETWYDITGNRSDTVKIGSDGWGEFHVNDGSVSIY

SEO ID No. 5

15

aaattcgatattgaaaacgattacaaataaaaattataatagacgtaaacgttcgagggt-20 ttgCtccctttttactCttt ttatgcaatcgtttcccttaatttttttggaagccaaaccgtcgaatgtaacatttgattaagggggaagggcatt

gtgct aacgtttcaccgcatcattcgaaaaggatggatgttcctgctcgcgtttttgctcactgtctcgctgttctgcccaacag gacagccgccaaggctGCCGCACCGT-25 TTAACGGCACCATGATGCAGTATTTTGAATGGTACTTGCCGGATGATGGCACG TTATGG-ACCAAAGTGGCCAATGAAGCCAACAACTTATCCAGCCTTGGCATCACCGCTCTTTGGCTG-CCGCCCGCTTACAA AGGAACAAGCCGCAGCGACGTAGGGTACGGAGTATACGACTTGTA-TGACCTCGGCGAATTCAATCAAAAAGGGACCGTCC GCACAAAATACGGAACAAAAGCTC-AATATCTTCAAGCCATTCAAGCCGCCCACGCCGCTGGAATGCAAGTGTACGCCGAT GTC-30 GTGTTCGACCATAAAGGCGGCGCTGACGGCACGGAATGGGTGGACGCCGTCGAAGTCAAT-CCGTCCGACCGCAACCA AGAAATCTCGGGCACCTATCAAATCCAAGCATGGACGAAATT-TGATTTTCCCGGGGGGGACACCCTACTCCAGCTTTA AGTGGCGCTGGTACCATTTTG-ACGGCGTTGATTGGGACGAAAGCCGAAAATTGAGCCGCATTTACAAATTCCGCGGCATC GGCAAAGCGTGGGATTGGGAAGTAGACACGGAAAACGGAAACTATGACTACTTAATGTAT -35 GCCGACCTTGATATGGATCA TCCCGAAGTCGTGACCGAGCTGAAAAACTGGGGGAAATG-GTATGTCAACACAACGAACATTGATGGGTTCCGGCTTGATG CCGTCAAGCATATTAAGT-TCAGTTTTTTCCTGATTGGTTGTCGTATGTGCGTTCTCAGACTGGCAAGCCGCTATTTACC GTCGGGGAATATTGGAGCTATGACATCAACAAGTTGCACAATTACATTACGAAAACAGAC-

87

GGAACGATGTCTTTGTTTGA TGCCCCGTTACACAACAAATTTTATACCGCTTCCAAATCAGGGGGCGCATTTGATATGCGCACGTTAATGACCAATACTC TCATGAAAGATCAACCGACATTGGCCGTCACCTTCGTTGATAATCATGACACCGGACCCGGCCAAGCGCTGCAGTCATGG GTCGACCCATGGTTCAAAACCGTTGGCTTTACGCTTTATTCTAACTCGG5 CAGGAAGGATACCCGTGCGTCTTTTATGGTGA CTATTATGCATTCACAAATATAACATTCCTTCGCTGAAAAACCAAAATCGATCCGCTCCTCATCGCGGCAGGGATTATG CTTACGGAACGCAACATGATATACTTGATCACTCCGACACTATCGGGTGGACAAGGGAAGGGACCAACCTGAAAAACCAGA TCCGGACTGGCCGCACTGATCACCGATGGGCCGGAGGAAGCAAATGGATGTACGTTGGCAAACACACGCTGGAAAAGT GTTCTATGACCTTACCGGCAACC-

10 GAGTGACACCGTCACCATCAACAGTGATGGATGGGGGGAATTCAAAGTCAATGGCG GTT-CGGTTTCGGTTTGGGTTCCTAGAAAAACGACCGTTTCTACCATCGCTCGGCCGATCACAA-CCCGACCGTGGACTGGT GAATTCGTCCGTTGGACCGAACCACGGTTGGTGGCATGGCCTTGA

tgcctgcga

15

SEO ID No. 6

AAPFNGTMMQYFEWYLPDDGTLWTKVANEANNLSSLGITA

LAULPPAYKGTSRSDVGYGYYDLYDLGBENQKGTVRTKYGT

KAQYLQAIQAAHAAGMQVYADVVFDHKGGADGTEWVDAVE
VNPSDRNQEISGTYQIQAWTKFDFPGRGNTYSSFKWRWYH
FDGVDWDESRKLSRIYKFRGIGKAWDMEVDTENGNYDYLM
YADLDMDHPEVVTELKNWGKWYVNTTNIDGFRLDAVKHIK
FSFFPDWLSYVRSQTGKPLFTVGEYWSYDINKLHNYITKT
DGTMSLFDAPLHNKFYTASKSGGAFDMRTLMTNTLMKDQP
TLAVTFVDNHDTEPGQALQSWVDFWFRPLAYAFILTRCBG
YPCVFYGDYYGIPQYNIPSLKSKIDPLLIARRDYAYGTQH
DYLDHSDIIGWTREGGTEKPGSGLAALITDGPGGSKWMYV

30 GKQHAGKVFYDLTGNRSDTVTINSDGWGEFKVNGGSVSVW VPRKTTVSTIARPITTRPWTGEFVRWTEPRLVAW

SEO ID No. 10

- 1 ATPADWRSOS IYFLLTDRFA RTDGSTTATC
 - 31 NTADQKYCGG TWQGIIDKLD YIQGMGFTAI
 - 61 WITPVTAOLP OTTAYGDAYH GYWOODIYSL
 - 91 NENYGTADDL KALSSALHER GMYLMVDVVA

	121	NHMGYDGAGS	SVDYSVFKPF	SSQDYFHPFC
	151	FIQNYEDQTQ	VEDCWLGDNT	VSLPDLDTTK
	181	DAAKNEMADM	VGSLVSNYSI	DGLRIDTVKH
	211	VQKDFWPGYN	KAAGVYCIGE	VLDGDPAYTC
5	241	${\tt PYQNVMDGVL}$	NYPIYYPLLN	AFKSTSGSMD
	271	DLYNMINTVK	SDCPDSTLLG	TFVENHONPR
	301	FASYTNDIAL	AKNVAAFIIL	NDGIPIIYAG
	331	QEQHYAGGND	PANREATWLS	${\tt GYPTDSELYK}$
	361	LIASANAIRN	YAISKDTGFV	TYKNWPIYKD
10	391	DITIAMRKGT	DGSQIVTILS	NKGASGDSYT
	421	LSLSGAGYTA	GQQLTEVIGC	TTVTVGSDGN
	451	VPVPMAGGLP	RVLYPTEKLA	GSKICSSS

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| March | Marc

| Colonia | Colo

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SUBSTITUTE SHEET (RULE 26)

Appendix 1

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| No. | 177 | Co. | 17.0 | Co.

Appendix 1 SUBSTITUTE SHEET (RULE 26)

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| No. | Control | Control

Appendix 1 SUBSTITUTE SHEET (RULE 26)

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| No. | No.

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| No. | Col. | C

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| No. | No.

SUBSTITUTE SHEET (RULE 26)

Appendix 1

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Appendix 1 SUBSTITUTE SHEET (RULE 26)

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Appendix 1 SUBSTITUTE SHEET (RULE 26)

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Appendix 1 SUBSTITUTE SHEET (RULE 26)

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| 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100

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Appendix 1 SUBSTITUTE SHEET (RULE 26)

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SUBSTITUTE SHEET (RULE 26)

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Appendix 1 SUBSTITUTE SHEET (RULE 26)

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SUBSTITUTE SHEET (RULE 26)

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| 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100

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SUBSTITUTE SHEET (RULE 26)

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121

MARCO MARCO

122

STATE CONTRACTOR OF CO

SUBSTITUTE SHEET (RULE 26)

2/27/2010, EAST Version: 2.4.1.1

123

| 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,000 | 1,00

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SUBSTITUTE SHEET (RULE 26)

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Appendix 1 SUBSTITUTE SHEET (RULE 26)

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CLAIMS

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- 1. A method of constructing a variant of a parent Termamyl-like α -amylase, which variant has α -amylase activity and at least sone altered property as compared to said parent α -amylase, which method comprises
- i) analysing the structure of the parent Termamyl-like α -amylase to identify at least one amino acid residue or at least 10 one structural part of the Termamyl-like α -amylase structure, which amino acid residue or structural part is believed to be of relevance for altering said property of the parent Termamyl-like α -amylase (as evaluated on the basis of structural or functional considerations),
- ii) constructing a Termamyl-like α -amylase variant, which as compared to the parent Termamyl-like α -amylase, has been modified in the amino acid residue or structural part identified in i) so as to alter said property, and
 - iii) testing the resulting Termamyl-like $\alpha\text{-amylase}$ variant for said property.
- 2. The method according to claim 1, wherein the property to be 25 altered is selected from the group consisting of substrate specificity, substrate binding, substrate cleavage pattern, temperature stability, pH dependent activity, pH dependent stability (especially increased stability at low (e.g. pH<6) or high (e.g. pH>9) pH values), stability towards oxidation, Ca²⁻¹ dependency and specific activity.
- 3. The method according to claim 1 or 2, wherein the property to be altered is the calcium ion dependency and the structural part to be modified is selected from the group consisting of 3s the C domain, the interface between the A and B domain, the interface between the A and C domain, or the interaction to a calcium binding site of the Termamyl-like α -amylase.

- 4. The method according to claim 1 or 2, wherein the property to be altered is the substrate cleavage pattern and the structural part to be modified is located within 10Å from an amino acid residue of the substrate binding site.
- 5. A method of constructing a variant of a parent Termamyl-like α -amylase, which variant has α -amylase activity and one or more altered properties as compared to said parent α -amylase, which method comprises
- 10 i) comparing the three-dimensional structure of the Termamyl-like α -amylase with the structure of a non-Termamyl-like α -amylase,
- ii) identifying a part of the Termamyl-like α -amylase structure which is different from the non-Termamyl-like α -amylase 15 structure and which from structural or functional considerations is contemplated to be responsible for differences in one or more properties of the Termamyl-like and non-Termamyl-like α -amylase, and
- iii) modifying the part of the Termamyl-like α -amylase 20 identified in ii) whereby a Termamyl-like α -amylase variant is obtained, one or more properties of which differ from the parent Termamyl-like α -amylase.
- 6. The method according to claim 6, wherein, in step iii), the 25 part of the Termamyl-like α-amylase is modified so as to ressemble the corresponding part of the non-Termamyl-like αamylase.
- 7. The method according to claim 5 or 6, wherein, in step iii), 30 the modification is accomplished by deleting one or more amino acid residues of the part of the Termamyl-like α -amylase to be modified; by replacing one or more amino acid residues of the part of the Termamyl-like α -amylase to be modified with the amino acid residues occupying corresponding positions in the 35 non-Termamyl-like α -amylase; or by insertion of one or more amino acid residues present in the non-Termamyl-like α -amylase into a corresponding position in the Termamyl-like α -amylase.

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- 8. The method according to any of claims 5-7, wherein the non-Termamyl-like α -amylase structure is the structure of a fungal α -amylase or a mammalian α -amylase.
- 5 9. The method according to claim 8, wherein the non-Termamyl-like α -amylase is the Aspergillus oryzae TAKA α -amylase, the A. niger acid α -amylase, the Bacillus subtilis α -amylase or the pig pancreatic α -amylase.
- 10 10. The method according to any of claims 1-9, wherein the parent Termamyl-like α -amylase is derived from a strain of Bacillus.
- 11. The method according to claim 10, wherein the parent α-15 amylase is derived from a strain of a B. licheniformis, B. amyloliquefaciens, B. stearothermophilus or a strain from an alkalophilic Bacillus sp. such as NCIB 12289, NCIB 12512 or NCIB 12513.
- 20 12. The method according to any of claims 1-11, wherein the parent α -amylase is a hybrid α -amylase comprising a combination of partial amino acid sequences derived from at least two α -amylases, of which one is a Termamyl-like α -amylase and the other(s) are, e.g., from a microbial and/or a mammalian α -25 amylase.
- 13. The method according to any of claims 5-12, wherein the part of the parent Termamyl-like α -amylase to be modified and identified in step ii) is loop 1, loop 2, loop 3 and/or loop 8 of the parent α -amylase.
 - 13. A method of constructing a variant of a parent Termamyllike α -amylase, which has a decreased calcium ion dependency as compared to said parent, which method comprises:
 - i) identifying an amino acid residue within 10Å from a Ca²-binding site of a Termamyl-like α -amylase in a model of the three-dimensional structure of said α -amylase, which from

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structural or functional considerations is believed to be responsible for a non-optimal calcium ion interaction.

- ii) constructing a variant in which said amino acid residue is replaced with another amino acid residue which from structural
 5 or functional considerations is believed to be important for establishing a higher Ca^{2*} binding affinity, and
 - iii) testing the Ca^{2^*} dependency of the resulting Termamyl-like α -amylase variant.
- 10 14. A method of constructing a variant of a parent Termamyllike α -amylase which variant has α -amylase activity and an altered pH dependent activity, which method comprises
- i) in a three-dimensional structure of the Termamyl-like α -15 amylase in question, identifying an amino acid residue within 15Å from an active site residue, in particular 10Å from an active site residue, which amino acid residue is contemplated to be involved in electrostatic or hydrophobic interactions with an active site residue.
- ii) replacing, in the structure, said amino acid residue with an amino acid residue which changes the electrostatic and/or hydrophobic surroundings of an active site residue and evaluating the accomodation of the amino acid residue in the 25 structure,
 - iii) optionally repeating step i) and/or ii) until an amino acid replacement has been identified which is accommodated into the structure,
 - iv) constructing a Termamyl-like α -amylase variant resulting from steps i), ii) and optionally iii) and testing the pH dependent activity of said variant.
- 35 15. A method of increasing the thermostability and/or altering the temperature optimum of a parent Termamyl-like α -amylase, which method comprises

- i) identifying an internal hole or a crevice of the parent Termamyl-like α -amylase in the three-dimensional structure of said α -amylase,
- ii) replacing, in the structure, one or more amino acid s residues in the neighbourhood of the hole or crevice identified in i) with another amino acid residue which from structural or functional considerations is believed to increase the hydrophobic interaction and to fill out or reduce the size of the hole or crevice.
- 10 iii) constructing a Termamyl-like α -amylase variant resulting from step ii) and testing the thermostability and/or temperature optimum of the variant.
- 16. A method of constructing a variant of a Termamyl-like α -15 amylase which has a reduced ability to cleave a substrate close to the branching point, which method comprises
- i) identifying the substrate binding area of the parent Termamyl-like α -amylase in a model of the three-dimensional structure of said α -amylase,
- ii) replacing, in the model, one or more amino acid residues of the substrate binding area of the cleft identified in i), which is/are believed to be responsible for the cleavage pattern of 5 the parent α-amylase, with another amino acid residue which from structural considerations is believed to result in an altered substrate cleavage pattern, or deleting one or more amino acid residues of the substrate binding area contemplated to introduce favourable interactions to the substrate or adding one or more amino acid residues to the substrate binding area contemplated to introduce favourable interactions to the substrate, and
- iii) constructing a Termamyl-like α -amylase variant resulting from step ii) and testing the substrate cleavage pattern of the 35 variant.
 - 17. The method according to any of the preceeding claims, in which the α -amylase variant is obtained by cultivating a

microorganism comprising a DNA sequence encoding the variant under conditions which are conducive for producing the variant, and optionally subsequently recovering the variant from the resulting culture broth.

- 18. A variant of a parent Termamyl-like α-amylase, in which variant at least one amino acid residue of the parent α-amylase, which is/are present in a fragment corresponding to the amino acid fragment 44-57 of the amino acid sequence of SEQ
 10 ID No. 4, has been deleted or replaced with one or more amino acid residues which is/are present in a fragment corresponding to the amino acid fragment 66-84 of the amino acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has been added using the relevant part of
 15 SEQ ID No. 10 or a corresponding part of another Fungamyl-like α-amylase as a template.
- 19. A variant of a parent Termamyl-like α -amylase, which variant has a region which, when the amino acid sequence of variant is aligned most closely with the amino acid sequence of the said parent α -amylase, occupies the same position as the portion from residue X to residue Y of SEQ ID No 4, the said region having at least 80% sequence homology with the part of SEQ ID No 10 extending from residue Z to residue V of SEQ ID No 25 10, wherein
 - X is the amino acid residue occupying position 44, 45, 46, 47 or 48 of SEQ ID No. 4,
 - Y is the amino acid residue occupying position 51, 52, 53, 54, 55, 56 or 57 of SEQ ID No. 4,
- 30 Z is the amino acid residue occupying position 66, 67, 68, 69 or 70 of SEQ ID No. 10, and
 - V is the amino acid residue occupying position 78, 79, 80, 81, 82, 83 or 84 of SEQ ID No. 10.
- 35 20. The variant according to claim 18 or 19, wherein X is the amino acid residue occupying position 48 and Y the amino acid residue occupying position 51 of SEQ ID NO 4 and Z is the amino

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acid residue occupying position 70 and V the amino acid residue occupying position 78 in SEQ ID No 10.

- 21. A variant of a parent Termamyl-like α -amylase, in which s variant at least one of the amino acid residues of the parent α -amylase, which is/are present in an amino acid fragment corresponding to the amino acid fragment 195-202 of the amino acid sequence of SEQ ID No. 4, has been deleted or replaced with one or more of the amino acid residues which is/are present in an amino acid fragment corresponding to the amino acid fragment 165-177 of the amino acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has been added using the relevant part of SEQ ID No. 10 or a corresponding part of another Fungamyl-like α -amylase is as a template.
- 22. A variant of a parent Termamyl-like α -amylase, which variant has a region which, when the amino acid sequence of variant is aligned most closely with the amino acid sequence of the said parent α -amylase, occupies the same position as the portion from residue X to residue Y of SEQ ID No 4, the said region having at least 80%, such as 90% sequence homology with the part of SEQ ID No 10 extending from residue Z to residue V of SEQ ID No 10, wherein
- 25 X is the amino acid occupying position 195 or 196 of SEQ ID No. 4,
 - Y is the amino acid residue occupying position 198, 199, 200, 201, or 202 of SEQ ID No. 4,
 - Z is the amino acid residue occupying position 165 or 166 of SEQ ID No. 10, and
- V is the amino acid residue occupying position 173, 174, 175, $_{\rm 35}$ 176 or 177 of SEQ ID No. 10.
 - 23. The variant according to claim 21 or 22, in which the amino acid fragment of the parent $\alpha\text{-amylase},$ which corresponds to

amino acid residues 196-198 of SEQ ID No. 4, has been replaced with the amino acid fragment corresponding to amino acid residues 166-173 of the amino acid sequence shown in SEQ ID No. 10.

- 5
- A variant of a parent Termamyl-like α-amylase, in which variant at least one of the amino acid residues of the parent α-amylase, which is/are present in a fragment corresponding to the amino acid fragment 117-185 of the amino acid sequence of
 SEQ ID No. 4, has/have been deleted or replaced with one or more of the amino acid residues, which is/are present in an amino acid fragment corresponding to the amino acid fragment 98-210 of the amino acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has been added which one or more additional amino acid residues has been added to using the relevant part of SEQ ID No. 10 or a corresponding part of another Fungamyl-like α-amylase as a template.
- 25. A variant of a parent Termamyl-like α -amylase, which variant has a region which, when the amino acid sequence of variant is aligned most closely with the amino acid sequence of the said parent α -amylase, occupies the same position as the portion from residue X to residue Y of SEQ ID No 4, the said region having at least 80%, such as at least 90% sequence homology with the part of SEQ ID No 10 extending from residue 25 Z to residue V of SEQ ID No 10, wherein
 - X is the amino acid occupying position 117, 118, 119, 120 or 121 of SEQ ID No. 4,
- $_{\rm 30}$ Y is the amino acid occupying position 181, 182, 183, 184 or 185 of SEQ ID No. 4,
 - Z is the amino acid occupying position 98, 99, 100, 101, 102 of SEQ ID No. 10, and
- 35
- V is the amino acid occupying position 206, 207, 208, 209 or 210 of SEQ ID No. 10.

26. The variant according to claim 24 or 25, in which an amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 121-181 of SEQ ID No. 4, has been replaced with the amino acid fragment corresponding to amino acid 5 residues 102-206 of the amino acid sequence shown in SEQ ID No. 10.

27. A variant of a parent Termamyl-like α -amylase, in which variant at least one of the amino acid residues of the parent 10 α -amylase, which is/are present in a fragment corresponding to the amino acid fragment 117-181 of the amino acid sequence of SEQ ID No. 4, has/have been deleted or replaced with one or more of the amino acid residues, which is/are present in an amino acid fragment corresponding to the amino acid fragment to 98-206 of the amino acid sequence shown in SEQ ID No. 10, or in which one or more additional amino acid residues has been added using the relevant part of SEQ ID No. 10 or a corresponding part of another Fungamyl-like α -amylase as a template.

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30 121 of SEO ID No. 4,

SEO ID No. 4,

28. A variant of a parent Termamyl-like α -amylase, which variant has a region which, when the amino acid sequence of variant is aligned most closely with the amino acid sequence of the said parent α -amylase, occupies the same position as the 2s portion from residue X to residue Y of SEQ ID No 4, the said region having at least 80%, such as at least 90% sequence homology with the part of SEQ ID No 10 extending from residue Z to residue V of SEQ ID No 10, wherein X is the amino acid occupying position 117, 118, 119, 120 or

Y is the amino acid occupying position 174, 175, 176 or 177 of

35 Z is the amino acid occupying position 98, 99, 100, 101, 102 of

s Z is the amino acid occupying position 98, 99, 100, 101, 102 of SEO ID No. 10, and

V is the amino acid occupying position 199, 200, 201 or 202 of SEQ ID No. 10.

- 29. The variant according to claim 27 or 28, in which the amino s acid fragment of the parent α -amylase, which corresponds to amino acid residues 121-174 of SEQ ID No. 4, has been replaced with the amino acid fragment corresponding to amino acid residues 102-199 of the amino acid sequence shown in SEQ ID No. 10.
- 30. A variant of a parent Termamyl-like α-amylase, in which variant at least one of the amino acid residues of the parent α-amylase, which is/are present in an amino acid fragment corresponding to the amino acid fragment 12-19 of the amino 15 acid sequence of SEQ ID No. 4, has/have been deleted or replaced with one or more of the amino acid residues, which is/are present in an amino acid fragment which corresponds to the amino acid fragment 28-42 of SEQ ID No. 10, or in which one or more additional amino acid residues has/have been inserted using the relevant part of SEQ ID No. 10 or a corresponding
- 31. A variant of a parent Termamyl-like α -amylase, which variant has a region which, when the amino acid sequence of 25 variant is aligned most closely with the amino acid sequence of the said parent α -amylase, occupies the same position as the portion from residue X to residue Y of SEQ ID No 4, the said region having at least 80%, such as at least 90% sequence homology with the part of SEQ ID No 10 extending from residue 30 Z to residue V of SEQ ID No 10, wherein

part of another Fungamyl-like α -amylase as a template.

- X is the amino acid occupying position 12, 13 or 14 of SEQ ID No. 4,
- Y is the amino acid occupying position 15, 16, 17, 18 or 19 of SEO ID No. 4.
- $^{\rm 35}$ Z is the amino acid occupying position 28, 29, 30, 31 or 32 of SEQ ID No. 10, and
 - V is an amino acid residue corresponding to the amino acid occupying position 38, 39, 40, 41 or 42 of SEO ID No. 10.

- 32. The variant according to claim 30 or 31, in which the amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 14-15 of SEQ ID No. 4, has been replaced with the amino acid fragment corresponding to amino acid 5 residues 32-38 of the amino acid sequence shown in SEQ ID No. 10.
- 33. A variant of a parent Termamyl-like α-amylase, in which variant at least one of the amino acid residues of the parent 10 α-amylase, which is present in a fragment corresponding to amino acid residues 7-23 of the amino acid sequence of SEQ ID No. 4, has/have been deleted or replaced with one or more amino acid residues, which is/are present in an amino acid fragment corresponding to amino acid residues 13-45 of the amino acid 15 sequence shown in SEQ ID No. 10, or or in which one or more additional amino acid residues has/have been inserted using the relevant part of SEQ ID No. 10 or a corresponding part of another Fungamyl-like α-amylase as a template.
- $_{20}$ 34. A variant of a parent Termamyl-like $\alpha\text{-amylase},$ which variant has a region which, when the amino acid sequence of variant is aligned most closely with the amino acid sequence of the said parent $\alpha\text{-amylase},$ occupies the same position as the portion from residue X to residue Y of SEQ ID No 4, the said $_{25}$ region having at least 80%, such as at least 90% sequence homology with the part of SEQ ID No 10 extending from residue Z to residue V of SEQ ID No 10, wherein
 - X is the amino acid occupying position 7 or 8 of SEQ ID No. 4,
- 30 Y is the amino acid occupying position 18, 19, 20, 21, 22 or 23 of SEQ ID No. 4,
 - Z is the amino acid occupying position 13 or 14 of SEQ ID No. 10, and
 - V is the amino acid occupying position 40, 41, 42, 43, 44 or 45 of SEO ID No. 10.

- 35. The variant according to claim 33 or 34, in which the amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 8-18 of SEQ ID No. 4, has been replaced with the amino acid fragment corresponding to amino acid 5 residues 14-40 of the amino acid sequence shown in SEQ ID No. 10.
- 36. A variant of a parent Termamyl-like α -amylase, in which variant at least one of the amino acid residues of the parent 10 α -amylase, which is present in a fragment corresponding to amino acid residues 322-346 of the amino acid sequence of SEQ ID No. 2, has/have been deleted or replaced with one or more amino acid residues, which is/are present in an amino acid fragment corresponding to amino acid residues 291-313 of the 15 amino acid sequence shown in SEQ ID No. 10, or or in which one or more additional amino acid residues has/have been inserted using the relevant part of SEQ ID No. 10 or a corresponding part of another Fungamyl-like α -amylase as a template.
- 20 37. A variant of a parent Termamyl-like α -amylase, which variant has a region which, when the amino acid sequence of variant is aligned most closely with the amino acid sequence of the said parent α -amylase, occupies the same position as the portion from residue X to residue Y of SEQ ID No 2, the said 25 region having at least 80% sequence homology with the part of SEQ ID No 10 extending from residue Z to residue V of SEQ ID No 10, wherein

X is the amino acid occupying position 322, 323, 324 or 325 of SEQ ID No. 2,

Y is the amino acid occupying position 343, 344, 345 or 346 of SEQ ID No. 2,

Z is the amino acid occupying position 291, 292, 293 or 294 of 35 SEQ ID No. 10, and

V is the amino acid occupying position 310, 311, 312 or 313 of SEQ ID No. 10.

38. The variant according to claim 36 or 37, in which the amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 325-345 of SEQ D No. 2, has been replaced with the amino acid fragment corresponding to amino acid residues 294-313 of the amino acid sequence shown in SEQ ID No. 10.

39. A variant of a parent Fungamyl-like α-amylase, in which variant at least one of the amino acid residues of the parent 10 α-amylase, which is/are present in an amino acid fragment corresponding to amino acid residues 291-313 of the amino acid sequence of SEQ ID No. 10, has/have been deleted or replaced with one or more of the amino acid residues, which is/are present in an amino acid fragment corresponding to amino acid residues 98-210 of the amino acid sequence shown in SEQ ID No. 4, or in which one or more additional amino acid residues has/have been inserted using the relevant part of SEQ ID No. 4 or a corresponding part of another Termamyl-like α-amylase as a template.

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40. A variant of a parent Fungamyl-like α -amylase, which variant has a region which, when the amino acid sequence of variant is aligned most closely with the amino acid sequence of the said parent α -amylase, occupies the same position as the 25 portion from residue X to residue Y of SEQ ID No 10, the said region having at least 80%, such as at least 90% sequence homology with the part of SEQ ID No 10 extending from residue Z to residue V of SEQ ID No 4, wherein

X is the amino acid occupying position 117, 118, 119, 120 or 30 121 of SEO ID No. 10,

Y is the amino acid occupying position 181, 182, 183, 184 or 185 of SEO ID No. 10,

35 Z is the amino acid occupying position 98, 99, 100, 101 or 102 of SEQ ID No. 4, and

V is the amino acid occupying position 206, 207, 208, 209 or 210 of SEQ ID No. 4.

41. The variant according to claim 39 or 40, in which the amino sacid fragment of the parent α -amylase, which corresponds to amino acid residues 121-181 of SEQ ID No. 10, has been replaced with the amino acid fragment corresponding to amino acid residues 102-206 of the amino acid sequence shown in SEQ ID No. 4.

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- 42. A variant according to any of claims 39-41, in which the the amino acid fragment of the parent α -amylase, which corresponds to amino acid residues 121-174 of SEQ ID No. 10, has been replaced with the amino acid fragment corresponding to amino acid residues 102-199 of the amino acid sequence shown in SEQ ID No. 4.
- 43. A variant of a parent Fungamyl-like α -amylase, in which an amino acid fragment corresponding to amino acid residues 181-20 184 of the amino acid sequence shown in SEQ ID No. 10 has been deleted.
- 45. A variant of a parent Termamyl-like α -amylase, which exhibits α -amylase activity and which has a decreased Ca^{2^*} dependency as compared to the parent α -amylase.
 - 46. A variant according to claim 45, which comprises a mutation in a position corresponding to at least one of the following positions in SEQ ID NO 2:
- 30 N104, A349, I479, L346, I430, N457, K385, F350, I411, H408 or G303, in particular a mutation corrsponding to N104D;

A349C+I479C;

L346C+I430C;

35 N457D,E;

N457D, E+K385R;

F350D, E+I430R, K;

F350D.E+I411R.K:

H408Q,E,N,D; and/or G303N,D,Q,E.

47. A variant of a parent Termamyl-like α -amylase which sexhibits a higher activity below the pH optimum than the parent α -amylase, which variant comprises a mutation of an amino acid residue corresponding to at least one of the following positions of the B. licheniformis α -amylase (SEQ ID NO 2): E336, Q333, P331, I236, V102, A232, I103, L196, in particular at least one of the following mutations:

E336R,K; Q333R,K; P331R,K; V102R,K,A,T,S,G; I236K,R,N; 15 I103K,R;

15 I103K,R; L196K,R; and/or A232T,S,G.

48. A variant of a parent Termamyl-like α -amylase which 20 exhibits a higher activity above the pH optimum than the parent α -amylase, which variant comprises a mutation of an amino acid residue corresponding to at least one of the following positions of the B. licheniformis α -amylase (SEQ ID NO 2): N236, H281 and/or Y273, in particular one of the following

25 mutations:
 N326I,Y,F,L,V;
 H281F,I,L; and/or
 Y273F.W.

30 49. A variant of a parent Termamyl-like α -amylase which exhibits α -amylase activity and which has an increased thermostability and/or altered temperature optimum as compared to the parent α -amylase, which variant comprises a mutation of an amino acid residue corresponding to at least one of the Solowing positions of the B. licheniformis α -amylase (SEQ ID NO 2):

```
L61, Y62, F67, K106, G145, I212, S151, R214, Y150, F143, R146,
   L241, I236, L7, V259, F284, F350, F343, L427 and/or V481, in
   particular at least one of the following mutations:
   L61W, V, F;
 5 Y62W;
   F67W:
   K106R, F, W;
   G145F, W
   I212F.L.W.Y.R.K:
10 S151 replaced with any other amino acid residue and in
   particular with F, W, I or L;
   R214W:
   Y150R.K:
   F143W:
15 R146W;
  L241I, F.Y.W:
   I236L, F, W, Y;
  L7F. I.W:
  V259F, I, L;
20 F284W;
  F350W:
  F343W:
  L427F, L, W; and/or
  V481, F, I, L, W.
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  50. A variant of a parent Termamyl-like q-amylase, which
  exhibits \alpha-amylase activity and which has a reduced capability
  of cleaving an oligo-saccharide substrate close to the
  branching point as compared to the parent \alpha-amylase, which
30 Variant comprises a mutation of an amino acid residue
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V54, D53, Y56, Q333 and/or G57, in particular at least one of 3s the following mutations:
V54L,I,F,Y,W,R,K,H,E,Q;

B. licheniformis α -amylase (SEQ ID NO 2):

corresponding to at least one of the following positions of the

D53L,I,F,Y,W;

Y.56W:

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0333W; and/or

G57 to all possible amino acid residues.

- 51. The variant according to any of claims 17-50, wherein one s or more proline residues present in the amino acid residues with which the parent α -amylase is modified are replaced with a non-proline residue such as alanine.
- 52. The variant according to any of claims 17-51, wherein one 10 or more cysteine residues present in the amino acid residues with which the parent α -amylase is modified are replaced with a non-cysteine residue such as alanine.
- 53. A DNA construct comprising a DNA sequence encoding an α -15 amylase variant according to any of claims 17-52.
 - 54. A recombinant expression vector which carries a DNA construct according to Claim 53.
- 20 55. A cell which is transformed with a DNA construct according to Claim 53 or a vector according to Claim 54.
 - 56. A cell according to Claim 55, which is a microorganism.
- 25 57. A cell according to Claim 56, which is a bacterium or a fungus.
- 58. The cell according to Claim 57, which is a grampositive bacterium such as Bacillus subtilis, Bacillus licheniformis, 30 Bacillus lentus, Bacillus brevis, Bacillus stearothermophilus, Bacillus alkalophilus, Bacillus amyloliquefaciens, Bacillus coagulans, Bacillus circulans, Bacillus lautus or Bacillus thuringiensis.
- 35 59. Use of an α -amylase variant according to any of claims 17-52 for washing and/or dishwashing.

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- 60. Use of an α -amylase variant according to any of claims 17-52 for desizing.
- 61. Use of an α -amylase variant according to any of claims 17-52 for starch liquefaction.
 - 62. A detergent additive comprising an α -amylase variant according to any of claims 17-52, optionally in the form of a non-dusting granulate, stabilised liquid or protected enzyme.
 - 63. A detergent additive according to Claim 62 which contains 0.02-200 mg of enzyme protein/g of the additive.
- 64. A detergent additive according to Claim 62 or 63, which is additionally comprises another enzyme such as a protease, a lipase, a peroxidase, another amylolytic enzyme and/or a cellulase.
- 65. A detergent composition comprising an α -amylase variant according to any of claims 17-52.
 - 66. A detergent composition according to Claim 65 which additionally comprises another enzyme such as a protease, a lipase, a peroxidase, another amylolytic enzyme and/or a cellulase.
 - 67. A manual or automatic dishwashing detergent composition comprising an α -amylase variant according to any of claims 17-52.
- 30 68. A dishwashing detergent composition according to Claim 67 which additionally comprises another enzyme such as a protease, a lipase, a peroxidase, another amylolytic enzyme and/or a cellulase.
- 35 69. A manual or automatic laundry washing composition comprising an α -amylase variant according to any of claims 17-52.

70. A laundry washing composition according to Claim 69, which additionally comprises another enzyme such as a protease, a lipase, a peroxidase, an amylolytic enzyme and/or a cellulase.

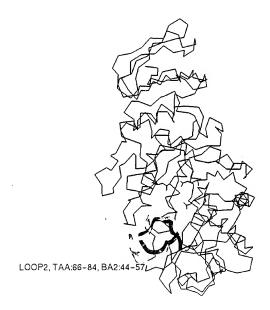


Fig. 1

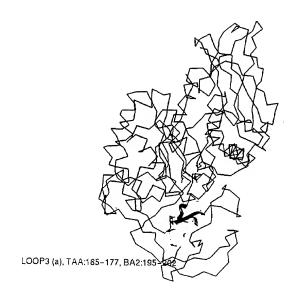


Fig. 2

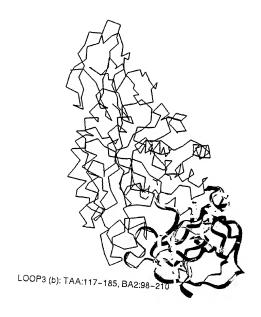


Fig. 3



Fig. 4

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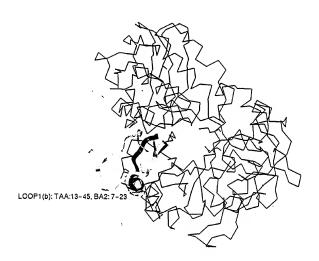


Fig. 5

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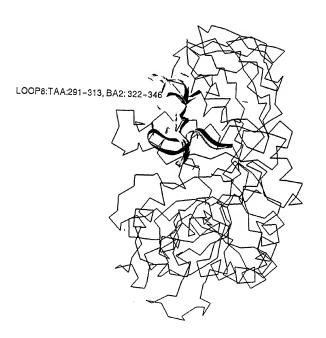


Fig. 6



Fig. 7

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CAT CAT AAT GGA ACA AAT GGT ACT ATG ATG CAA TAT TTC GAA TGG TAT TTG CCA AAT GAC

н	н	N	G	T	N	G	T	M	M	Q	Y	F	E	w	Y	Ľ	P	n An	D	
21																				
G	N	H	M	N AAC	R R	L	R	D	D	GC/	l GCT	r AAC	TT.	A AAG	AG!	r aa	A GG	G AT	A ACA	
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41	CTA	TCC.					-													
A	V	W	I	P	P	A	W	K	G	T	r TCC	CAC	AA:	r GAT	GT/	GG*	r TA	r GG	GCC	
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61		-		~~																
Y	D	L	Y	D	L	GGA	E GAG	F	AAC N	CAG	i AAG	GGG	ACC	GTI	CGI	, YC	· AA	TA:	r GGA	
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81																				
T	R	N	Q.	L	O	A	A A	V	ACC T	S	TTA	. AAA	. AAT	. AAC	GGC	ATT	CAC	GTA	TAT	
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101		-	-	N.T.C																
G	D	V	V	M	N	H	K	GGT	GGA	GCA A	GAT D	GGT	ACG	GAA	ATT	GTA	AAT	, GCC	GTA	
												•	•	-	•	•	.,	^	•	
121	GTG	327	ccc	NCC	***	CCA	***	~~	~~											
E	v	N.	R	5	N	R	N N	Q	E	T	S	GGA	GAG	TAT	GCA	ATA	GAA	. GCG	TGG W	
														-		•	-			
141	AAG	بلحثملة	CAT	بالحلحك	CCT	CCA	ACA.	ccs	337		C . T		٠							
T	ĸ	F	D	F	P	G	R	G	N	N	н	s	S	F	AAG K	W	R	TGG W	Y	
161																				
	TTT	GAT	GGG	ACA	GAT	TGG	GAT	CAG	TC2	ccc	CAC		CAR	240		3 Th	Th T		mac	
н	F	D	G	T	D	w	D	Q	s	R	Q	L	Q	N	ĸ	I	Y	X	F	
181																				
	GGA	ACA	GGC	AAG	GCC	TGG	GAC	TGG	GAA	GTC	GAT	ACA	GAG	AAT	GGC	440	TAT	GAC	тат	
R	G	T	G	ĸ	A	W	D	W	E	v	D	T	E	N	G	N	Y	D	Y	
201																				
	ATG	TAT	GCA	GAC	GTG	GAT	ATG	GAT	CAC	CCA	GAA	GTA	ATA	CAT	GAA	CTT	AGA	AAC	TGG	
	и																	N		
221																				
	GTG	TGG	TAT	ACG	AAT	ACA	CTG	AAC	стт	GAT	GGA	TTT	AGA	ATA	GAT	GCA	GTG	AAA	CAT	
	V											F								
241																				
ATA	AAA											GTG	CGT	AAC	ACC	ACA	CCT	AAA	CCA	
I	к	Y	S	F	т	R	D	W	L	T	н	v	R	N	т	Ŧ	G	К	>	

K T S W N H S V F D V P L H Y N L Y N A Fig. 8

ATG TTT GCA GTG GCT GAG TTT TGG AAA AAT GAC CTT GGT GCA ATT GAA AAC TAT TTG AAT M F A V A E F W K N D L G A I E N Y L N AAA ACA AGT TGG AAT CAC TCG GTG TTT GAT GTT CCT CTC CAC TAT AAT TTG TAC AAT GCA

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TOT AAT AGO GGT GGT TAT TAT GAT ATG AGA AAT ATT TTA AAT GGT TOT GTG CTG CAA AAA
S N S G G Y Y D M R N I L N G S V V Q K .
CAT CCA ACA CAT GCC GTT ACT TIT GTT GAT AAC CAT GAT TCT CAG CCC GGG GAA GCA TTG
H P T H A V T F V D N H D S Q P G E A L
GAA TCC TIT GIT CAA CAA TGG TIT AAA CCA CIT GCA TAT GCA TIG GIT CIG ACA AGG GAA
E S F V Q Q W F K P L A Y A L V L T R E
CAA GGT TAT CCT TCC GTA TTT TAT GGG GAT TAC TAC GGT ATC CCA ACC CAT GGT GTT CCG
OGYPSVFYGDYYGIPTHGVP
GCT ATG ANA TCT ANA ATA GAC CCT CTT CTG CAG GCA CGT CAA ACT TTT GCC TAT GGT ACG
AHKSKIDPLLQARQTFAYGT
CAG CAT GAT TAC TTT GAT CAT CAT GAT ATT ATC GGT TGG ACA AGA GAG GGA AAT AGC TCC
Q H D Y F D H H D I I G W T R E G N S S
CAT CCA AAT TCA GGC CTT GCC ACC ATT ATG TCA GAT GGT CCA GGT GGT AAC AAA TGG ATG
H P N S G L A T I M S D G P G G N K W
TAT GTG GGG AAA AAT AAA GCG GGA CAA GTT TGG AGA GAT ATT ACC GGA AAT AGG ACA GGC
Y V G K N K Å G Q V W R D I T G N R T G
ACC GTC ACA ATT AAT GCA GAC GGA TGG GGT AAT TTC TCT GTT AAT GGA GGG TCC GTT TCG
T V T I N A D G W G N F S V N G G S V S
481
GTT TGG GTG AAG CAA TAA
v w v k Q ·
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Fig. 8 (cont.)

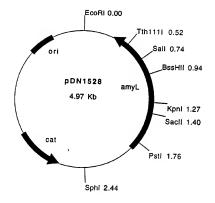


Fig. 9

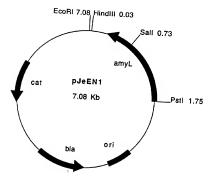


Fig. 10

International application No. PCT/DK 96/00057

A. CLASSIFICATION OF SUBJECT MATTER

IPC6: C12N 9/28, C12N 15/56
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC6: C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

WPI, CA, MEDLINE, BIOSIS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	Dialog Information Services, File 5, BIOSIS PREVIEWS, Dialog accession no. 11619266, BIOSIS no. 98219266, Machius M et al: "Crystal structure of calcium-depleted Bacillus licheni- formis alpha-amylase at 2.2 A resolution", & Journal of Molecular Biology 246 (4). 1995. 545-559	1-17
х	Dialog Information Services, file 155, MEDLINE, Dialog accession no. 08974640, MEDLINE accession no. 94289640, Svensson B: "Protein engineering in the alpha-amylase family: catalytic mechanism, substrate specificity, and stability", & Plant Mol Biol (NETHERLANDS) May 1994, 25 (2) p141-57	1-17

х	Further documents are listed in the continuation of Box C.	X See patent family annex
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Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance

"E" eriter document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance: the claimed invention cannot be considered novel or cannot be coondered to involve an inventive step when the document is taken alone

"Y" document of particular relevance: the claimed invention caonol be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report 0.5 -07-1996
5 July 1996	
Name and mailing address of the ISA	Authorized officer
Swedish Patent Office	
Box 5055, S-102 42 STOCKHOLM	Yvonne Siösteen
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Form PCT/ISA/210 (second sheet) (July 1992)

INVITATION TO PAY ADDITIONAL FEES

The invention claimed relates to a method of constructing alpha-amylase variants with predetermined properties by comparing the three-dimensional structures of enzymes. The claims also include many alpha-amylase variants.

"A search for a special technical feature" as mentioned in PCT Rule 13.2 among the independent claims did not reveal a . unifving, novel technical feature.

Accordingly, the following inventions were found:

- I Claims 1-17 focus on a method of constructing alphaamylase variants by comparing the tree-dimensional
 structure of a parent enzyme (Temamyl-like alpha-amylase)
 with another enzyme e.g. mammalie or fungal alphaamylases. The differences in structure are compared with
 the differences in function, whereafter new variants with
 new predictable characteristics are produced.
- II Claims 45-46 directed to a alpha-amylase variant that has decreased Ca2+ dependency,
- III Claim 47 directed to a alpha-amylase variant that exhibits higher activity below the ph-optimum than the parent enzyme.
- IV Claim 48 directed to a alpha-amylase variant having an increased thermostability and/or altered temperature optimum.
- V Claim 50 directed to a variant having reduced capability of cleaving an oligo-saccharide substrate close to its branching point.

Due to the complex construction of the claims and the fact that the search so far has not covered all aspects of the invention, it may be that further non-unity remarks can appear. If further searches are done, references might appear which will give furter a posterior; non-unity remarks.

Therefore, the search has been restricted to the first invention.

Form PCT/ISA/206 (extra sheet) (July 1992)

International application No.

PCT/DK 96/00057

Claims 18-43 are directed to a number of different variants that are composed of several inventions. They are, however, so complex and broad that no meaningful search can be done, especially as no special characteristic is linked to the groups of variants. It is for example unlikely that claim 18 concerns one invention. It is not believable that a change in any amino acid in one fragment for one/or none of the amino acids in a fragment of another enzyme gives an enzyme with the same new and valuable characteristic. The formulation of claims 18-43 is so complicated because of all the different combinations of amino acid substitutions.

Thus they do not comply with Art. 6. PCT prescribing that claims shall be clear and concise.

Form PCT/ISA/210 (extra sheet) (July 1992)

International application No.
PCT/DK 96/00057

		96/00057
C (Continu	uation). DOCUMENTS CONSIDERED TO BE RELEVANT	
Category*	Citation of document, with indication, where appropriate, of the relevant passag	es Relevant to claim No
x	Dialog Information Services, file 155, MEDLINE. Dialog accession no. 08958150, MEDLINE accession no. 94273150, Nakatani H et al: "Effect of modify histidine residues on the action of Bacillus amylu liquefaciens and barley-malt alpha-amylases", & Carbohydr Res (NETHERLANDS) Apr 16 1994, 257 (1) p 155-61	p-
Y		45-46
х	J. MED. BIOL., Volume 229, 1993, C. Chang et al, "Crystallization and Preliminary X-ray Crystallographic Analysis of alpha-Amylase from Bacillus subtilis" page 235 - page 238	1-17
		
A	WO 9100343 A2 (GIST-BROCADES N.V.), 10 January 1991 (10.01.91)	1-17
٩	EP 0410498 A2 (GIST-BROCADES N.V.), 30 January 1991 (30.01.91)	1-17
		
^	JOURNAL OF BACTERIOLOGY, Volume 166, No 2, May 1986, G. L. Gray et al, "Structural Genes Encoding the Thermophilic alpha-Amylases of Bacillus stearothermophilus and Bacillus licheniformis" page 635 - page 643	1-17
		
Р, Х	WO 9535382 A2 (GISTBROCADES B.V.), 28 December 1995 (28.12.95), claims 1-2, abstract	45-46
	WO 9418314 A1 (GENENCOR INTERNATIONAL), 18 August 1994 (18.08.94)	45-46

International application No. PCT/DK 96/00057

	1075K 307K	
C (Continu	ation). DOCUMENTS CONSIDERED TO BE RELEVANT	
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim N
Y	Chemical Abstracts, Volume 108, No 11, 14 March 1988 (14.03.88), (Columbus, Ohio, USA), Buisson, G. et al., "Three dimensional structure of porcine pancreatic alpha-amylase at 2.9 Å resolution. Role of calcium in structure and activity", page 325, THE ABSTRACT No 90927h, EMBO J. 1987, 6 (13), 3909-3916	45-46
Y	Chemical Abstracts, Volume 112, No 15, 9 April 1990 (09.04.90), (Columbus, Ohio, USA), Vihinen, Mauno et al, "Site-directed mutagenesis of a thermostable alpha-amylase from Bacillus stearothermophilus: putative role of three conserved residues", page 347, IHE ABSTRACT No 135178r, J. Biochem 1990, 107 (2), 267-272	45-46
A	US 4600693 A (KAREN L. KINDLE ET AL), 15 July 1986 (15.07.86)	45-46
A	Chemical Abstracts, Volume 112, No 19, 7 May 1990 (07.05.90), (Columbus, Ohio, USA), Holm, Liisa et al, "Random mutagenesis used to probe the structure and function of Bacillus stearothermophilus alpha-amylase", page 351, THE ABSTRACT No 174785f, Protein Eng. 1990, 3 (3), 181-191	45-46
	· 	
	<u>.</u>	
	·	

International application No. PCT/DK96/00057

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
nos required to be searched by this Authority, namely:
2. X Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out second to the prescribed requirements to such
an extent that no meaningful international application that do not comply with the prescribed requirements to such
see next sheet
3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a). Box II Observations where unity of learning the second and third sentences of Rule 6.4(a).
and the state of invention is tacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
see next sheet
STATE STILLED
As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims for
Claims 1-17 directed to a method of constructing alpha-amylase variants and claims 45-46 directed to an alpha-amylase.
No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
, his device by Gains 1405.:
Remark on Protest The additional count G
Remark on Protest The additional search fees were accompanied by the applicant's protest. X No protest accompanied the payment of additional search fees.
personnel search lees.

Form PCTASA 210 (continuation of first sheet (1)) (July 1992)

INTERNATIONAL SEARCH REPORT Information on patent family members

International application No.
01/04/96 PCT/DK 96/00057

	document arch report	Publication date	Patent family member(s)			Publication date	
IO-A2-	9100343	10/01/91	AU-B,B- AU-A-	629959 5939790 2032518		15/10/92 17/01/91	
			CA-A- EP-A,A,A JP-T-		99	30/12/90 23/01/91 06/02/92	
D-A2-	0410498	30/01/91	AU-B- AU-A- CA-A- CN-A- JP-T- US-A- WO-A,A,A	6382 59538 20305 10502 45007 53647 91003	90 54 20 56 82	24/06/93 17/01/91 30/12/90 27/03/91 13/02/92 15/11/94 10/01/91	
-A2-	9535382	28/12/95	NONE				
)-A1-	9418314	18/08/94	NONE				
5-A-	4600693	15/07/86	NONE				

Form PCT/ISA.210 (patent family annex) (July 1992)